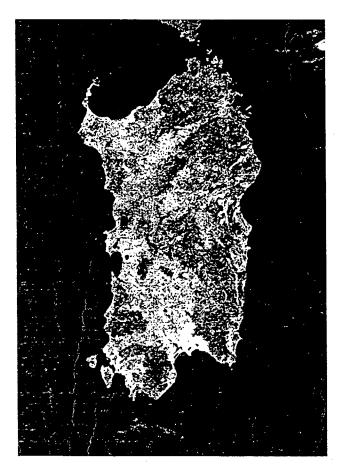
5th Conference on Recombinant Protein Production

A comparative view on host physiology

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ABSTRACT BOOK

Stable expression of a human-like sialylated recombinant thyrotropin in a chinese hamster ovary cell line expressing $\alpha 2$,6-sialyltransferase

Bartolini P¹, Oliveira JE¹, Damiani R¹, Vorauer-Uhl K², Peroni CN¹, <u>Ribela MTCP¹</u>

¹Biotechnology Department, IPEN-CNEN, University of São Paulo, Brazil (email:mtribela@ipen.br) ²Department of Biotechnology, University of Natural Resources and Applied Life Sciences, Vienna, Austria

Recombinant thyrotropin (r-hTSH), expressed in CHO cells, lacks α 2,6-sialic acid-galactose linkages at the termini of the carbohydrate chains, unlike pituitaryderived hTSH (p-hTSH) that contains approximately 70% of α 2,3- and 30% of α 2,6- linked sialic acid residues. In this work, a genetic modification in the hTSHproducing CHO cell line by the introduction of rat α 2,6-sialyltransferase cDNA was carried out, generating a human-like r-hTSH (hlr-hTSH) more similar to the native hormone, with respectively 61 ± 10 % of α 2,3- and 39 ± 10 % of α 2,6linked sialic acid residues. The best clone, isolated from this "human-like" CHO cell line co-transfected with the dicistronic expression vectors pEDdc- α and pEDdc- β hTSH and submitted to gene amplification with up to 8 µM methotrexate (MTX), presented a secretion level of 2.1 µg hTSH/10⁶ cells/per day, which is useful for production purification and characterization.

The relative molecular masses (Mr) of the heterodimer and of the α - and β subunits of purified hlr-hTSH, obtained by MALDI-TOF mass spectrometry, were 29187, 14038 and 15243 respectively, which are in good agreement with previously determined values for r-hTSH. The carbohydrate structures of this rhTSH were of the complex type, presenting 57.1% of di-, 18.8% of tri- and 23.2% of tetra-antennary structures, partly fucosylated and with variable levels of sialvlation. The most abundant structures were the monosialvlated biantennary N-linked sugar chains, representing ~ 45% of all identified forms, followed by triand tetra-sialylated N-linked sugar chains (~17.5% each). About 86% of oligosaccharides were siglylated, with 5.47 moles siglic acid/mole protein, being the sialic acid: galactose ratio = 0.73. "Human-like" r-hTSH, analyzed via an in vivo bioassay based on hTSH-induced T₄ release, was shown to be equipotent (p> 0.05) with the unique commercial preparation of r-hTSH (Thyrogen), and 1.6fold more potent than p-hTSH-NIDDK(p<0.001). We are showing below an example of sialic acid linkage analysis for p-hTSH and hlr-hTSH based on specific lectin interaction.

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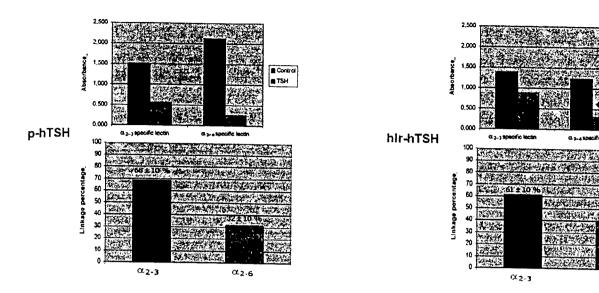
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