



Natural factors and chemical contamination control the structure of macrobenthic communities in the Santos Estuarine System (SP, Brazil)

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Keywords: Benthos; Distribution; Macrofauna; Pollution; Santos Estuarine System.

Abstract: The Santos Estuarine System (SES) is a complex of bays, islands, estuarine channels, and rivers located on the Southeast coast of Brazil, in which multiple contaminant sources are situated in close proximity to mangroves and other protected areas. In this study, the composition and structure of the macrobenthic communities of SES were described and ninety-nine species were identified, with the predominance of polychaetes and bivalve mollusks. The benthic assemblages also showed strong signs of stress, as indicated by the low abundance, richness and diversity, and the dominance of opportunistic species. Integrated analysis including sediment characteristics related to natural and anthropogenic factors (e.g., sediment chemistry, and toxicity) indicated that benthic fauna from the inner portions of the SES and vicinities of the SSOS diffusers as Santos Bay were affected. Some locations at the mouths of Santos and São Vicente estuaries exhibited moderate disturbance. In other sites from the mouth of São Vicente and Bertiooga channels, and Santos Bay, the benthic fauna were considered not degraded. Our results suggest that a combination of both environmental factors and contaminants were responsible for the benthic community structure.

Abbreviations: AH – aliphatic hydrocarbon; ANOVA – Analysis of variance; CEC – Compounds of emerging concern; DO – Dissolved oxygen; LOE – Lines of evidence; PAH – polycyclic aromatic hydrocarbon; MBAS – Methylene Blue Active Substance; nMDS – Non-metric multidimensional scaling; NOEC – No effective concentration; PCA – principal component analysis; PEL – Probable Effect Level; QA/QC – Quality Assurance and Quality Control procedure; SES – Santos Estuarine System; SSOS – Santos Sewage Outfall System; SQT – Sediment quality triad; TEL – Threshold Effect Levels; TOC – total organic carbon.

Nomenclature: Day (1967), Amaral (1980), Nonato (1981), Amaral and Nonato (1982, 1984), Lana (1984), Bolívar and Lana (1986), Amaral and Nonato (1994, 1996) for polychaetes, Rios (1984) for mollusks, Young (1998) for crustaceans, Borges et al (2002) for ophiuroids, and Migotto (1996) for cnidarians.

Introduction

Marine and estuarine ecosystems are influenced by a combination of physical, chemical and biological drivers that determine the structure of their communities (Reiss et al. 2015). Classic papers have reported that natural variables are the primary factors responsible for the structure of benthic communities (Ankar and Jansson 1973, Pearson and Rosenberg 1987), especially in physically controlled environments, as estuaries and other transitional waters (Tommasi 1979). Variables as salinity, temperature, dissolved oxygen, grain size, content of organic matter, among others, influence the distribution of macrobenthic organisms in coastal and estuarine waters (Grebmeier et al. 1989, Ozolin'sh 2000, Rosenberg et al. 2001). Anthropogenic factors such as pol-

lution may also have an influence on benthic communities, affecting thus the species composition and distribution (Reish 1986, 1993, Pearson and Rosenberg 1987, Dauvin and Ruellet 2009, Elliott and Quitino 2007, Muniz et al. 2008, Venturini et al. 2008).

The Santos Estuarine System (SES) is located on the central coast of São Paulo state, Brazil (23°30' - 24°00'S; 46°05' - 46°30'W) and it was once considered one of the world's most polluted estuaries (CETESB 1985, Tommasi 1979) due to impacts of multiple contamination sources, such as the Port of Santos, the industrial complex, the unplanned urbanization, increased domestic and industrial wastes dumping, and the uncontrolled expansion of tourism. During the 1990s, an important pollution abatement program was conducted by

the State Environmental Agency, which reduced the inputs of contaminants into the ecosystems (Lamparelli et al. 2001). However, the applied measures were not enough to restore the environmental quality of SES as demonstrated by several studies (Abessa et al. 1998, 2005, Cesar et al. 2006, Medeiros and Bicego, 2004, Martins et al. 2008, 2010, 2011, Pereira et al. 2011, 2014).

To date, sediments from the SES still present elevated concentrations of several chemicals at toxic levels for benthic organisms (Torres et al. 2009, Pereira et al. 2016, Santos et al. 2018) and some authors have shown that benthic macrofaunal assemblages are altered (Tommasi 1979, Abessa et al. 2008, Buruaem et al. 2013). These studies highlighted that pollution caused the changes, but they could not discern definitively if the alterations were due to natural factors, contaminants or a combination of both. In estuaries, disentangling the effects on benthic communities caused by natural factors from those induced by contamination is challenging because these areas are subjected to periodic variations of salinity, tides, organic matter input, currents and waves (Elliott and Quintino 2007, Dauvin and Ruellet 2009).

Integrated approaches using multiple lines of evidence (LOEs) have been recommended to deal with these challenges (Long and Chapman 1985, Burton Jr et al. 2002, Cesar et al. 2007) and they were also applied in SES. In particular, the Sediment Quality Triad (SQT) explores the relationships between sediment chemistry and laboratory-based toxicity assays with the ecological indicators of macrobenthic communities (Chapman 1990, Long et al. 2000, Burton Jr et al. 2002; Chapman and Hollert 2006, Zamboni and Abessa 2002). However, the specific composition of benthic organisms and the influence of natural factors are not considered in such methods and these data are relevant as baseline information for ecological studies and environmental management. Thus, the present study aimed to primarily characterize the macrobenthic community from the SES and evaluate the

main factors responsible for its distribution along this estuarine system. We focused on identifying the roles of natural and anthropogenic factors on benthic organisms in order to fill the gaps generated in the environmental quality studies and provide information for management and conservation programs.

Materials and methods

Sediment sampling

Sediment and biological samples were collected from 20th to 23rd March 1998, at 25 sampling sites distributed along the SES using a stainless steel Petersen grab sampler, with 0.026 m² of the sampling area (Fig. 1). Sites were chosen based on the occurrence of natural gradients at Santos (St. 1 to 6) and São Vicente estuaries (St. 9 to 13), and also considering the presence of multiple contamination sources from the Industrial complex at Cubatão (St. 7 and 8) and from the sewage effluent discharges at the Santos Bay (St. 14 to 23). Two sites were located at Bertioga channel in order to cover the whole extent of the estuarine system (St. 24 and 25). Sites were georeferenced using a Garmin 38 GPS, and the geographical coordinates are shown in the supplementary material (Table S1).

For each site, the surficial 2-cm layer of the retained sediments was collected and samples from approximately 10 replicates were pooled for the analyses of physical-chemical parameters and toxicity bioassays. Sub-samples were separated and frozen until their use for grain-size distribution, total organic carbon (TOC) and physical-chemical analyses, while another batch for toxicity tests were kept refrigerated at 4 ± 2 °C for 3-5 days. The sediment pore water was also extracted by the suction method (Winger and Lasier 1991), using 100 ml syringes, and then centrifuged for 20 minutes at 4200 g

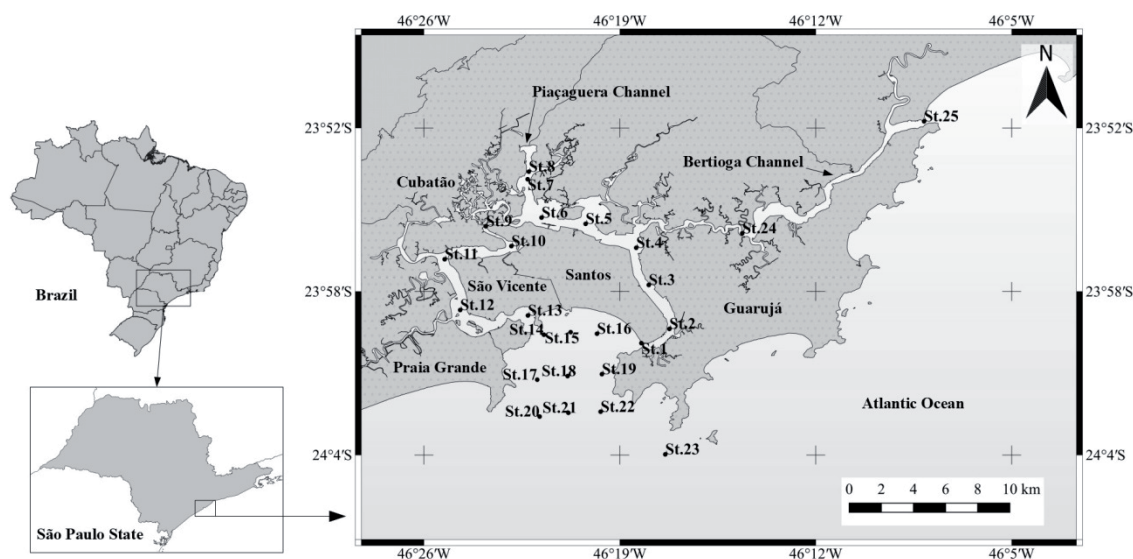


Figure 1. Map of the Santos Estuarine System - SES (SP, Brazil) showing the sampling stations.

(Carr et al. 1996, 2000, 2001), at 4 °C. The supernatant was transferred to amber glass flasks and stored frozen.

For the macrofauna, three independent replicates were obtained from the 5-10 cm layer of sediment. The collected material was sieved in a 0.5 mm mesh, and the biological material retained fixed in 4% formaldehyde for 24 h and then it was transferred to 70% alcohol. In each site, bottom water samples were collected using Nansen bottles (i.e., at less than 1 meter from the bottom), and their physical-chemical properties were measured in the field (See Table S2, Supplementary Material). The salinity was measured using an ATAGO-S/Mill refractometer (0.5% precision); the dissolved oxygen content was measured with a TOA oximeter (DO-14P, 0.01 mg/L precision); pH values were estimated by using Macherey-Nagel colorimetric indicator sticks (0.1 precision); and the water temperature was measured with digital thermometers (0.1 °C precision).

Sediment properties

The grain size distribution was analyzed by the wet sieving method to separate fine fraction (silt + clay), followed by dry sieving to separate gravel and sand fractions (Suguio 1973, McCave and Syvitski 1991). The calcium carbonate contents were estimated according to the acid digestion method (Gross, 1971), which consists of 10% HCl digestion followed by the gravimetry of samples. Decarbonated sediments were analyzed for the contents of total organic carbon (TOC), nitrogen (N) and sulfur (S) using an automated analyzer model LECO CNS 2000 and the Micro Kjeldahl method (McKenzie and Wallace 1954).

The concentrations of Al, Fe, Cd, Cr, Co, Hg, Ni, Pb and Zn were analyzed in sediment extracts. A microwaves system in high pressure was used for the total digestion of Al, Cr, and Fe (CEM Corporation, model MDS – 2000) in a solution composed by Milli-Q water, HNO₃, HF, and HCl. For Cd, Co, Pb, and Zn, a similar procedure was employed, but using a solution of aqua regia and HClO₄ in the extraction. Elements were detected using a fast sequential Atomic Absorption Spectroscopy. For Hg, samples were extracted in aqua regia and HClO₄ heated for 30 min at 90 °C (Akagi and Nishimura 1991). The extracts were introduced into a system of Flux Injection for Cold Vapour generation (FIA-CV-AAS) (Fostier et al. 1995) and analyzed with an Atomic Absorption Spectroscopy (VARIAN, AAS 220-FS).

Sediments were also analyzed for aliphatic hydrocarbons (AHs) and polycyclic aromatic hydrocarbons (PAHs), according to the protocol described by UNEP (1991). An amount of 25 g was Soxhlet-extracted with a 50% mixture of n-hexane and dichloromethane for 8 hours and the extracts were fractionated into F1 (AHs) and F2 (PAHs) by silica gel-alumina column chromatography. Then, samples were eluted with n-hexane (F1) and 30% dichloromethane in n-hexane (F2). Twenty-six AHs (n-C17 to n-C35, including pristine and phytane) were analyzed in a Gas Chromatographer (GC, Hewlett Packard, model 5890 II) equipped with the flame ionization detector maintained at 325 °C (Table S3, Supplementary

Material). Twenty-six PAHs were analyzed in a GC, coupled to a mass spectrometer in the Ion Monitoring System mode (V.G. Masslab – Fisons, model TRIO 1000) (Table S4, Supplementary Material). Concentrations were determined by the analyses of calibration curves, and the method detection limit (MDL) was calculated as 3 times the standard deviation from 7 replicates of a sample which contained surrogates. MDL values were 0.07 µg.g⁻¹, for the total n-alkanes, and 0.24 ng.g⁻¹ for the ΣPAHs. The detection limits for individual compounds ranged from 0.0002 to 0.0112 µg.g⁻¹ for n-alkanes and from 0.002 to 0.022 ng.g⁻¹ for the ΣPAHs.

Surfactants concentrations in sediments were estimated by the Methylene Blue Active Substance (MBAS) method (APHA/AWWA/WEF 1998) after extraction by elutriation in distilled water (Abessa and Sousa 2004). The procedure involved three successive extractions from acid aqueous medium containing methylene blue into chloroform, followed by an aqueous backwash and measurement of the blue color in the chloroform by spectrophotometry at 652 nm. As for Quality Assurance and Quality Control procedure (QA/QC), methods for determinations were validated by the analysis of certified reference sediment Buffalo River® (for metals) and NIST (for hydrocarbons). The concentrations found for metals and PAHs were compared with Canadian sediment quality guidelines (Smith et al. 1996, Environment Canada 1999), which have been used as an environmental quality benchmark by the São Paulo State Environmental Agency (Lamparelli et al. 2001).

Toxicity tests

The whole sediment toxicity tests with the amphipod *Tiburonella viscana* followed the protocol described by Melo and Abessa (2002) and ABNT (2016). Sediment samples were thoroughly homogenized, and aliquots of approximately 250 ml were introduced into the test chambers (1-L polyethylene flasks) containing 750 ml of filtered seawater. Ten amphipods were added to each test chamber. The system was prepared in three replicates per sample and incubated for 10 days at 25 ± 2 °C, under constant aeration and light. After the exposure period, samples were sieved through a 0.5-mm screen and the surviving amphipods were counted, with organisms considered dead. The effects induced by samples and reference sediments (control) were compared by Student t'-test (Zar 1984), and sample considered toxic when the amphipod survival was statistically lower than that exhibited by the reference group (p < 0.05). Levels of dissolved oxygen, salinity and pH of the overlying water in the test chambers were measured at the beginning and end of the test.

Toxicity of porewater samples were evaluated by the early life stage bioassay with embryos of the sea urchin *Lytechinus variegatus* (CETESB 1992; USEPA 1988). Prior to the test, samples were thawed at 25 °C and analyzed for salinity, pH, dissolved oxygen and temperature. The total ammonia concentration was measured by a colorimetric method (Koroleff 1970) and the unionized ammonia levels were estimated according to Whitfield (1974) model. Adult individuals of *L. variegatus* were collected at rocky reefs in Ubatuba,

and transported to the laboratory. The spawning was induced by the injection of KCl (0.5 M) into the celomatic cavities of the animals and the fertilization was performed *in vitro*. The exposure of approximately 500 embryos to the samples was initiated using glass tubes as test chamber containing 10 ml of porewater from the samples. Four replicates were prepared for each treatment, which included 3 porewater dilutions: 100, 50 and 10% (Carr et al. 2001). The experiment was incubated for 24h at 25 ± 2 °C. After the exposure, the test was finished by the addition of 10% buffered formaldehyde into each replicate. The embryos were analyzed microscopically for morphological anomalies and retarded development (100 per replicate). The results were statistically analyzed by analysis of variance (ANOVA), followed by the Dunnett's *t* test comparison using the SAS statistical package (SAS 1992).

Macrobenthic community

The biological material was identified to the lowest possible taxonomic level (i.e., to species when possible) by using identification keys, specifically for polychaetes (Day 1967, Amaral 1980, Nonato 1981, Amaral and Nonato 1982, 1984, Lana 1984, Bolívar and Lana 1986, Amaral and Nonato 1994, 1996), and mollusks (Rios 1984). Voucher samples for other groups were identified by experts. The results obtained for each sampling station were expressed as the mean number of organisms collected in a 0.026 m². The density of individuals was expressed in ind/m² in order to allow comparisons to other studies. For each sampling site, the Species Richness, Shannon Diversity, Simpson Dominance, and Polychaetes Density (Magurran 1998) were estimated by using the software package Bio Diversity Professional (Lambshhead et al. 1997).

Species frequency (F) was also calculated for each taxon by the following index (Guille 1970): $F = (P_p / P) \times 100$, where: P_p is the number of stations where the taxon occurred and P is the total number of stations. Then, taxa were classified as constant (Cn; when $F > 50\%$), common (Co; when $10\% < F < 49\%$) and rare (Ra; when $F < 10\%$). Rare species were excluded and Bray–Curtis similarity matrices were constructed for both benthic macrofauna and environmental variables. A fourth root transformation was applied to the data to reduce the influence of abundant species. Non-metric multidimensional scaling (nMDS) ordinations were generated to investigate patterns in the structure of the macrofaunal assemblages.

Data integration

Different methods have been used to integrate complex data, as biological variables and their association to the abiotic factors (Chapman et al. 1991, Green 1993, Green and Montagna 1996). In this study we used the principal component analysis (PCA), which consisted in the simultaneous ordination of two matrices in the same multidimensional space, one containing the physical-chemical variables and the other with biological data (species densities) (Del Valls and Chapman 1998, Del Valls et al. 1998). The first matrix includ-

ed variables of bottom water column (pH, OD, temperature, salinity) and sediment (depth, % of mud, TOC, CaCO₃, N, S, metals, hydrocarbons, surfactants and toxicity data). The second matrix was constructed with the mean density values of each benthic species. Data expressed in percentages were transformed using an arcsine function and then both matrices were transformed using a logarithmic function $\text{Log}(x + 1)$ in order to reduce differences in the scale of the variables. The cut-off value of |0.50| for factor loading was considered as a significant correlation to a respective component (Comrey and Lee 1992), in order to show the associations between abiotic variables and species distribution.

Results and discussion

The physical and chemical parameters of water column are presented in the Supplementary Material (Table S2). The salinities of surface waters were lower in the inner parts of the estuary, while the salinities at the bottom were higher, corroborating the saline intrusion within the system (Bonetti 2000, Tommasi 1979) and also confirming the freshwater drainage from the continental basins, which flows through the surface waters (Harari et al. 2000). Water temperatures were slightly higher at the surface compared to the bottom, reflecting summer conditions in this subtropical estuary. The pH values were typical from marine/estuarine waters, with higher values observed at the Santos Bay and lower values reported within the estuary and close to the sewage outfall diffusers (St. 18) as reported by Rachid (2002). The dissolved oxygen levels in the bottom water ranged from 3.0 to 7.2 mg O₂/L. The lowest values occurred in the inner parts of Santos and Bertioga Channels (stations 4, 5, 6, 7, 8 and 24). Levels of dissolved oxygen were saturated with lower values reported in the inner parts of Santos and Bertioga Channels (St. 4 to 8, and 24). In general, water characteristics were consistent with descriptions of the SES provided by following studies, with continental inputs influencing the estuarine channels and marine waters acting mainly at the Bay (Bonetti 2000, Moser 2002).

The sediments of SES presented a variable composition in terms of grain-size and levels of TOC, N, and S (Table 1). Sites from Santos and Bertioga Channel exhibited muddy sediments. At São Vicente Channel, the mouth of Bertioga Channel, and west side of Santos Bay sediments were sandy. At the central and east portions of Santos Bay, which are influenced by discharges of the Santos Sewage Outfall System (SSOS) and inputs from Santos Channel, mixtures of mud and sands were found. The contents of CaCO₃ were higher in the central portion of the bay and the inner part of the Santos Channel. Elevated levels of TOC, N and S occurred at high concentrations in estuarine zones and close to the SSOS outfalls at Santos Bay, along with muddy sediments, indicating the occurrence of the depositional areas, as reported in the literature (Fúlfaro and Ponçano 1976, Fúlfaro et al. 1983, Tommasi 1979, Fukumoto et al. 2004). Sedimentation rates in the Santos Channel are low, with most of the particles retained in the upper estuary with impacts of human interven-

Table 1. Sediment grain-size and organic characteristics in the SES and reference areas.

Station	% Sand	% Mud	%CaCO ₃	TOC (%)	S (%)	N (%)
St. 1	14.82	85.18	11.20	1.39	0.55	0.12
St. 2	6.40	93.60	21.20	2.53	0.43	0.27
St. 3	0.80	99.20	21.20	2.37	0.64	0.22
St. 4	29.70	80.73	23.90	1.03	0.34	0.09
St. 5	2.12	97.88	20.30	2.14	0.62	0.16
St. 6	8.82	91.18	11.90	0.79	0.13	0.07
St. 7	11.46	88.54	11.20	1.39	0.55	0.12
St. 8	56.41	43.59	14.17	2.76	0.41	0.27
St. 9	60.60	39.40	11.50	2.62	0.41	0.11
St. 10	93.33	6.77	12.10	2.03	0.12	0.09
St. 11	91.16	8.84	11.20	2.51	0.49	0.08
St. 12	98.33	1.77	6.38	0.31	0.01	0
St. 13	98.34	1.76	11.33	1.22	0.02	0.01
St. 14	97.03	2.97	5.20	0.12	0.01	0
St. 15	95.73	4.27	6.29	0.14	0.01	0
St. 16	72.74	27.26	11.33	0.70	0.13	0.06
St. 17	92.17	7.83	7.43	0.23	0.03	0
St. 18	28.84	71.16	21.66	1.39	1.43	0.12
St. 19	33.26	66.74	17.30	1.55	0.67	0.14
St. 20	97.43	2.57	5.29	0.14	0.01	0
St. 21	41.97	58.03	21.27	1.17	0.55	0.09
St. 22	88.03	11.97	8.35	0.29	0.04	0.02
St. 23	45.53	54.47	7.70	0.21	0.03	0.05
St. 24	9.82	90.18	11.30	0.87	0.09	0.06
St. 25	98.74	1.21	4.73	0.77	0.05	0.07

Sand = particles > 0.0625 mm; Mud = particles ≤ 0.0625 (silts + clays).

tions affecting such processes in the adjacent areas (Fúlfaro and Ponçano 1976, Fúlfaro et al. 1983).

The concentrations of metals in sediments from the SES are shown in Table 2. For most of elements analyzed, the highest concentrations tended to occur in the sediments from the inner portion of the SES. A detailed discussion of these results is presented in Abessa et al. (2008).

The higher concentrations of Al and Fe occurred in sediments from the upper estuary, with a decreasing gradient towards the sea. These data corroborate the results of previous studies (Bonetti 2000; Luiz-Silva et al. 2006) and also indicate inputs from industrial or terrestrial origins since a major steel plant is located at Piacaguera channel. Elevated concentrations of trace elements Cd, Cr, Hg, Ni, Pb, and Zn were found above threshold values at the inner portions of Piacaguera, Santos, and Bertioga channels (St. 4 to 9, and St. 24). The concentrations of Co were high in sediments from the upper estuary and moderate levels close to the sewage outfalls and at the West part of the Santos Bay. These levels are in accordance with other studies and can be explained by the presence of multiple sources of contamination such as port activities, the industrial complex, urban runoff and sewage releases (Prósperi et al. 1998, Lamparelli et al. 2001, Bonetti 2000, Luiz-Silva et al. 2006). Recent studies showed that the pattern of distribution reported in our study remains unchanged (Bordon et al. 2011, 2018, Kim et al. 2016, 2017). The contamination of sediments by metals is relevant in the

SES, because these chemicals are toxic, can bioaccumulate and be transferred through the trophic chain, producing ecological effects.

The results of n-alkanes, PAHs and MBAS are shown in Table 3. Details of these analyses, including the concentrations of each compound analyzed are displayed in the Supplementary Material (Tables S3 and S4) and discussed in Abessa (2002). Levels of n-alkanes occurred systematically in SES, with higher concentrations observed in estuarine zones and the SSOS vicinity, at Santos Bay (Table 3). The presence of such compounds is a proxy of AHs contamination (Medeiros and Bicego 2004). In most samples, n-alkanes were mainly from biogenic sources, according to odd/even compounds and pristane/phytane relationships, as expected in an estuarine system surrounded by mangroves (Nishigima et al. 2001). In the sediments from port and industrial areas (St. 1 to 8, 10, 18 and 25), the contribution of petrogenic hydrocarbons was also relevant.

The distribution of PAHs was restricted to the Santos Channel (Table 3). The concentrations of some individual PAHs were also elevated, above threshold values (Probable Effect Levels - PELs and/or Threshold Effect Levels - TELs). A decreasing gradient toward the lower estuary was also observed, suggesting that the sources of PAHs are located in the upper estuarine portion. Our results are in accordance with the other studies (Medeiros and Bicego 2004, Bonetti 2000, Bicego et al. 2006). Concentrations reported here

Table 2. Concentrations of metals (in % and µg/g) in sediments from the Santos Estuarine System. Numbers and italic and light-grey cells indicate values above the TEL; numbers in bold and dark-grey cells indicate values above the PEL.

Stations	Metals								
	Al	Fe	Zn	Ni	Pb	Cd	Cr	Co	Hg
	%		µg/g						
St. 1	3.04	1.21	40.1	9.5	10.9	< 0.50	18.7	6.0	0.11
St. 2	3.00	0.87	47.6	8.9	11.2	< 0.50	17.6	5.2	0.12
St. 3	3.11	0.92	44.5	7.0	10.8	< 0.50	7.5	4.2	<i>0.36</i>
St. 4	5.34	2.77	<i>180.0</i>	<i>21.8</i>	204.8	<i>0.75</i>	37.9	10.7	0.74
St. 5	5.21	2.68	284.4	22.2	23.5	<i>0.92</i>	44.1	10.3	<i>0.23</i>
St. 6	6.07	2.75	86.9	<i>25.0</i>	19.2	<i>0.99</i>	44.8	12.3	<i>0.32</i>
St. 7	7.91	4.99	<i>152.8</i>	<i>34.1</i>	<i>39.7</i>	0.42	<i>65.8</i>	17.0	0.92
St. 8	6.19	7.99	312.0	44.2	<i>89.9</i>	<i>0.98</i>	<i>97.5</i>	15.3	0.75
St. 9	3.33	1.76	<i>77.6</i>	13.2	19.6	<i>1.49</i>	22.8	5.1	<i>0.50</i>
St. 10	0.58	0.63	14.2	2.5	3.7	< 0.50	5.0	0.9	0.11
St. 11	5.91	2.85	37.9	10.2	10.3	< 0.50	<i>53.6</i>	4.8	<i>0.31</i>
St. 12	0.08	0.52	7.6	1.3	2.5	< 0.50	5.0	0.2	0.04
St. 13	0.06	0.62	10.9	2.4	17.0	< 0.50	5.0	1.1	<0.03
St. 14	3.42	1.27	34.0	9.1	6.5	< 0.50	12.5	4.1	0.05
St. 15	2.71	1.77	41.4	11.3	8.3	< 0.50	18.8	5.8	0.04
St. 16	1.64	0.83	23.8	4.9	5.3	< 0.50	10.0	2.3	0.04
St. 17	2.96	1.25	35.9	10.3	7.8	< 0.50	18.4	5.4	0.04
St. 18	3.56	1.60	61.7	12.5	16.8	< 0.50	28.4	7.1	<i>0.19</i>
St. 19	3.75	1.70	44.7	13.4	11.8	< 0.50	29.0	6.5	0.06
St. 20	2.70	1.11	49.6	7.9	5.25	< 0.50	9.5	3.2	0.04
St. 21	2.60	1.23	32.2	14.7	5.75	< 0.50	19.6	4.1	0.04
St. 22	4.53	2.07	55.5	<i>17.9</i>	18	< 0.50	40.9	8.5	0.076
St. 23	2.28	1.14	29.7	8.1	5.5	< 0.50	5.0	3.8	<0.030
St. 24	5.53	2.35	74.4	<i>21.2</i>	24.5	<i>0.85</i>	<i>69.5</i>	11.6	0.11
St. 25	1.00	0.61	16.8	5.9	<2.0	< 0.50	5.0	1.6	0.037
TEL	-	-	<i>124.0</i>	<i>15.9</i>	<i>30.2</i>	<i>0.70</i>	<i>52.3</i>	-	<i>0.130</i>
PEL	-	-	271.0	42.8	122.0	4.21	160.0	-	0.696

were higher than those found in São Sebastião (Weber et al. 1998) and similar to those reported to the Montevideo Harbor (Muniz et al. 2004) and Todos os Santos Bay (Venturini and Tommasi 2004). Levels of detergents (MBAS) were detected close to the SSOS diffusers, followed by the stations from São Vicente channel, corroborating the ranges observed by Tommasi (1979).

Results of whole sediment toxicity to *T. viscana* are presented in the Fig. 2. The samples from the estuarine channels (St. 1 to 10, 24 and 25) were considered toxic, as well as those from the stations 17 and 18, located close to the SSOS diffusers (St. 17 and 18) (Figure 2). The physical-chemical parameters of the overlying water within the test-chambers were considered suitable for *T. viscana* (Supplementary Material, Table S5). The results of the porewater toxicity test are shown in the Figure 3 and in Supplementary Materials (Tables S6 and S7). Samples from stations 20 and 25 were not tested. All the porewater samples were toxic and produced significant reduction of the larval development (excepting the St. 17 at 50%). The toxicity persisted in samples at 25% porewater dilution, with exception of the samples from the St. 10, 15, 16, 17 and 21. The unionized ammonia can contribute to the toxicity in porewater samples (Carr et al. 2001), and the no effective concentration (NOEC) of NH₃ for *L. variegatus* embryonic development is 0.05 mg/L (Prósperi 2002). In

this investigation, the toxicity occurred even when the NH₃ levels were low, suggesting that other contaminants were probably responsible for the toxicity. Some recent publications reported that ammonia is an important contaminant to the SES and is capable of interacting with other substances, increasing the toxic effects to benthic organisms (Araújo et al. 2013, Camargo et al. 2015, Moreira et al. 2019). A qualitative interpretation of the ammonia influence on the porewater toxicity is presented on the Supplementary Material (Table S7).

The stations from the estuarine zones and the SSOS vicinities were the most toxic. Despite the influence of contaminants released by port and industrial activities, recent studies also showed that the vicinities of the SSOS present contamination by contaminants of emerging concern (CECs), such as phenols, hormones, fecal sterols, pharmaceutical and personal care products, drugs of abuse, and others (Santos et al. 2018, Pereira et al. 2016). Our results are in agreement with previous studies carried out in the SES (Abessa et al. 2001, 2005, Abessa and Sousa 2001), and after our sampling campaign, other studies corroborated our findings (Sousa et al. 2007, Cesar et al. 2006, Rachid, 2002, Prósperi et al. 1998, Araújo et al. 2013, Buruaem et al. 2013, Moreira et al. 2019).

For macrobenthic community a total of 1,413 individuals were collected in SES. The raw data for benthos is available in the Supplementary Material (Table S8) and a detailed

Table 3. Concentrations of poly-aromatic, n-alkane hydrocarbons and detergents (estimated as Methylene Blue Active Substances – MBAS) in sediments from the Santos Estuarine System (µg/g).

Compounds (µg/g)	Station																									
	St. 1	St. 2	St. 3	St. 4	St. 5	St. 6	St. 7	St. 8	St. 9	St. 10	St. 11	St. 12	St. 13	St. 14	St. 15	St. 16	St. 17	St. 18	St. 19	St. 20	St. 21	St. 22	St. 23	St. 24	St. 25	
Total n-alkanes	1.05	1.83	1.48	2.12	3.06	2.87	3.71	4.09	1.77	4.29	9.31	0.02	0.03	1.4	0.49	0.53	1.03	4.09	2.08	0.31	1.29	1.58	0.88	3.21	0.4	
Pristane/Phytane	-	1.01	0.23	0.5	0.57	-	0.66	0.57	-	0.4	0.25	-	-	1	-	1	1	1.05	1	-	2	1	1	0.05	0.4	
odd/even	1.81	1.42	2.03	2.53	4.61	3.75	1.65	1.49	4.42	1.24	5.81	NC	2	3.69	4.33	3.9	3.59	1.67	4.69	3	3.54	3.61	3.72	4.37	1.46	
odd/even <23	1	1.42	2.28	1.18	1.36	1.74	1.27	1.88	1.8	1.76	1.52	NC	1	1.5	1.5	1.5	1.5	1.49	1.7	1.25	1.83	1.25	2.5	1.67	1	
odd/even >24	2.13	1.43	1.96	3.38	5.53	3.97	1.74	1.44	4.89	1.13	6.83	NC	NC	4.26	6.6	5.5	4.79	1.74	5.85	4.75	4	4.36	4.07	5.3	2.5	
UCM	NM	NM	NM	NM	NM	NM	NM	NM	NM	4.32	NM	54.4	-	0.59	10.2	3.2	4.52	8.82	72	21.2	2.59	7.51	15	6.21	15.3	8.18
Sum of PAHs*	0.08	0.19	2.91	1.68	39.8	28.8	11	42.4	2.12	1.38	0.03	0.002	0	0.02	0.01	0	0.01	0.51	0.01	0	0.01	0.01	0	0.03	0	
MBAS (µg/g)	0.98	2.93	1.47	NM	3.42	5.37	3.42	2.93	2.44	7.33	4.89	2.44	11.24	3.91	7.82	1.47	NM	13.68	NM	6.84	4.40	6.35	4.89	5.37	1.95	

NC = Not calculable; NM = not measured; UCM = Unresolved Complex Mixture

*Sum of 30 PAHs.

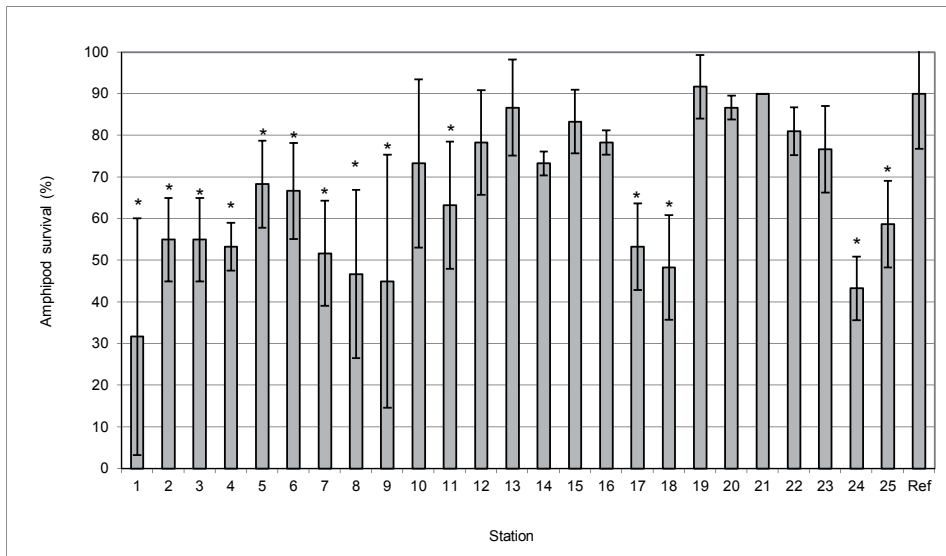


Figure 2. Whole sediment toxicity to the amphipod *T. viscana*, (* = significant difference ($p < 0.05$)).

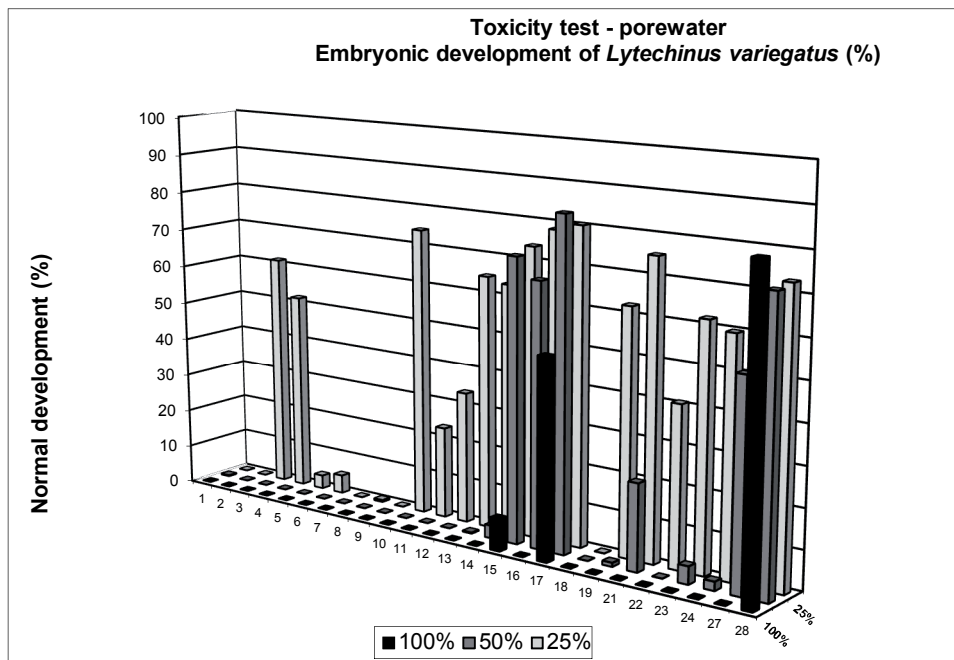


Figure 3. Mean normal embryonic development of *Lytechinus variegatus* exposed to pore water extracted from sediments from the Santos Estuarine System.

analysis including a “station by station” approach was provided in Abessa (2002). Samples exhibited few species, with predominance of polychaetes, followed by bivalve mollusks. The dominance of polychaetes was above 60%, whereas crustaceans occurred only at the stations 1, 13, 17 and 19. About 56 taxa of polychaetes were identified, 18 bivalves, 6 gastropods, 11 crustaceans, 2 ophiuroids, among others (see Table 4 for the list of species identified). The St. 16 presented the higher number of species (37 in total) and St. 22 exhibited the highest abundance, estimated in 10,564 individuals per m². Abundance was also higher at ST. 1, 16, 23, 24, and 25. No

organisms were found at St. 5 and 12, and low abundances were estimated at St. 4, 8, 10, 13, 14, 15, 17 and 20 (< 100 indiv./m²).

Most of the species found were considered rare, occurring at only one, two or three stations. The more common taxa were polychaetes, especially *Capitella capitata* and *Owenia fusiformis*. Other common taxa included the polychaetes *Rhodine* sp., *Magelona posterolongata*, *Ninoe brasiliensis*, *Ophioglycera* sp., *Hemipodus* sp., *Nephtys* sp., *Lumbrineris* sp. and *Diopatra cuprea*, the mollusks *Anachis obesa* (gastropod), *Chione cancelata*, *Tagellus* sp. and *Ctena pectinella*

Table 4. List of macrobenthic species identified in the Santos Estuarine System.

Taxa		
Polychaeta	Polychaeta	Penaeidae
<i>Rhodine sp</i>	<i>Spiophanes sp</i>	<i>Trachypenaeus constrictus</i>
<i>Magelona posterolongata</i>	<i>Eunice (sensu stricto)</i>	Thalassinidea
Goniadidae (unidentified)	<i>Marphysa sp</i>	<i>Upogebia affinis</i>
<i>Glycinde sp</i>	<i>Lysidice sp</i>	Paguroidea
<i>Goniadides sp</i>	<i>Paraeulepis sp</i>	<i>Clibanarius vitatus</i>
<i>Goniada sp</i>	<i>Cirrophorus sp</i>	Brachyura
<i>Ophioglycera sp</i>	<i>Paraonis sp</i>	<i>Tetraxanthus rathbunae</i>
Glyceridae (unidentified)	<i>Euphrosine sp</i>	<i>Cyrtoplax spinidentata</i>
<i>Hemipodus sp</i>	Hesionidae (unidentified)	Amphipoda
<i>Glycera sp</i>	Pilargidae (unidentified)	<i>Tiburonella viscana</i>
<i>Euzonus (Thoracophelia)</i>	Phylodocidae (unidentified)	<i>Photis longicaudata</i>
Nephtyidae (unidentified)	Sabellidae (unidentified)	Isopoda
<i>Nephtys sp.</i>	Gastropoda	<i>Excirollana armata</i>
Capitellidae (unidentified)	<i>Heleobia australis</i>	Tanaidacea
<i>Mediomastus sp.</i>	<i>Calyptrea centralis</i>	<i>Kalliapseudes schubartii</i>
<i>Heteromastus filiformis</i>	<i>Nassarius vibex</i>	Mysidacea
<i>Capitella capitata</i>	<i>Turbonilla nivea</i>	<i>Promysis atlantica</i>
<i>Dasybranchus sp</i>	<i>Anachis sertulariarum</i>	Dendrobranchiata (n.i.)
Terebellidae (prob. <i>Pista sp.</i>)	<i>Anachis obesa</i>	Teleostei
<i>Trichobranthus sp</i>	Nudibranchia	<i>Ribeiroclinus eigenmanni</i>
<i>Scoloplos (Leodamas) sp</i>	<i>Doris bovena</i>	Anthozoa
<i>Haploscoloplos sp</i>	<i>Etidoris ladislavii</i>	<i>Renila</i>
<i>Laeonereis acuta</i>	Bivalvia	<i>Edwardsia sp</i>
Nereidae (unidentified)	<i>Tellina sp</i>	Ophiuroidea
<i>Naineris setosa</i>	<i>Tellina alternata</i>	<i>Microphiopholis atra</i>
Prob. <i>Pseudeurythoe</i>	<i>Semele sp1</i>	<i>Ophionereis reticulata</i>
<i>Pectinaria sp</i>	<i>Semele purpurascens</i>	Nemertina (unidentified)
Dorvilleidae (unidentified)	<i>Ctena pectinella</i>	
Syllidae (unidentified)	<i>Felaniella candeana</i>	
<i>Typosilis sp</i>	<i>Periplona ovata</i>	
<i>Ninoe brasiliensis</i>	<i>Chione cancelata</i>	
<i>Lumbrineris sp</i>	<i>Crassinela martinicensis</i>	
Serpulidae (unidentified)	<i>Tagellus sp</i>	
<i>Owenia fusiformis</i>	<i>Mactra fragilis</i>	
Onuphidae (unidentified)	<i>Corbula patagonica</i>	
<i>Nothria</i>	<i>Strigilla carnaria</i>	
<i>Rhamphobranchium sp.</i>	<i>Laevicardium brasilianum</i>	
<i>Americanuphis</i>	<i>Ervilia sp2</i>	
<i>Diopatra cuprea</i>	<i>Tivella mactroides</i>	
Chaetopteridae (unidentified)	<i>Abra lioica</i>	
<i>Mesochaetopterus sp</i>	<i>Anomalocardia brasiliana</i>	
<i>Poecilochaetus sp</i>		
<i>Scolelepis sp</i>		
<i>Laonice sp.</i>		

(bivalves), and the ophiuroid *Microphiopholis atra*. Most of these species are typically from coastal marine regions or cosmopolitan, except for *Anachis obesa*, which is found in estuarine areas. Among these species, *C. capitata*, *O. fusiformis*, *M. posterolongata*, and *Nephtys sp.* are considered pollution tolerant, suggesting that the contamination influences on the benthic species composition. In general, the species richness was low, as expected for complex estuarine systems that are subject to extreme natural environmental variations and gradients (Table 5). Species richness was higher at stations lo-

cated on the east side of Santos Bay (St. 16, 19, 22 and 23), in the mouth of Santos Channel (St. 1 and 2) and in Bertioga Channel (St. 24). In Santos Bay, the west portion tended to exhibit few species (stations 14, 17 and 20) compared to the east side (St. 16, 19, 22 and 23).

Such pattern had been described previously by Tommasi (1979), but the specific composition of assemblages differed from those observed in the present study. A further study conducted by Moreira et al. (2019) confirmed the trends ob-

Table 5. Ecological Indices (Specific Richness; Simpson Dominance, Shannon Diversity (H') and polychaete abundances, in each sampling station.

Station	Ecological Indices			Polychaetes abundance (%)
	Specific Richness	Diversity Shannon	Simpson dominance	
St. 1	16	1.611	0.399	58.54
St. 2	15	2.380	0.134	37.93
St. 3	9	1.942	0.257	80.00
St. 4	1	0	1.000	0
St. 5	0	0	nc	0
St. 6	5	1.560	0.211	75.00
St. 7	10	1.033	0.610	94.91
St. 8	4	1.330	0.268	83.33
St. 9	3	0.656	0.641	100
St. 10	2	0.693	0.480	100
St. 11	8	1.774	0.204	92.86
St. 12	0	0	nc	nc
St. 13	1	0	1.000	0
St. 14	3	1.099	0.316	66.67
St. 15	4	1.332	0.269	80.00
St. 16	37	3.362	0.047	50.77
St. 17	3	1.040	0.363	0
St. 18	6	0.911	0.560	91.30
St. 19	14	2.432	0.104	88.00
St. 20	2	0.693	0.480	50.00
St. 21	9	1.946	0.190	85.71
St. 22	18	0.815	0.673	99.16
St. 23	18	2.642	0.088	85.71
St. 24	17	2.383	0.127	83.05
St. 25	13	2.108	0.154	97.32

nc = not calculable.

served at the west of Santos Bay, with dominance of polychaetes (Cirratulidae and Capitellidae) and presence of sensitive organisms such as amphipods. The pattern exhibited by Shannon diversity (H') was similar to species richness and values were low compared to a previously investigation (Tommasi, 1979), which can be attributed to differences in sampling strategy. The occurrence of few individuals and rare species in the majority of stations resulted in low values of Simpson dominance. The elevated dominance of polychaetes is also an indicator of altered communities typically found in naturally stressed environments as in the case of estuarine systems (Reish 1986, Van Dolah et al. 1999, Weisberg et al. 1997). The SES is a physically controlled environment, known to be complex and dynamic, where the abiotic variables are more important than interspecific relationships as controlling factors of benthic communities structures (Tommasi 1979). In this situation, the opportunistic organisms (r-strategists) are favored, as demonstrated by the opportunistic species, such as *Capitella capitata*, *Magelona posterolongata*, *Owenia fusiformis*, *Rhodine* sp, *Anachis obesa*, and *Chione cancelata*.

The use of nMDS analysis aimed to explore the influence of abiotic factors on the benthic fauna and the results for environmental variables (Fig. 4) showed that stations from Santos estuary towards Piaçaguera Channel (St. 1 to 9), were

dissimilar from the other stations, including Bertioga channel (St. 24, and 25), São Vicente estuary (St. 10-13) and Santos Bay (St. 14 to 23). On the other hand, such a pattern was not apparent in the structure of assemblages. However, there was a trend showing that some samples from Santos estuary (St 1, 3, 4, 6, 7 and 9) separated from the other stations in a similar manner, indicating a possible combination of multiple variables in the distribution of the species. The influence of natural stressors makes it difficult to discern the effects of contaminants on the benthic community structure of the SES. In disturbed sites, the disappearance of sensitive species followed by the dominance of resistant organisms (Gesteira and Dauvin 2000) is described in the literature as well the occurrence of lower densities and fewer species (Dumbauld et al. 2001, Lardicci et al. 2001).

Studies on benthic community ecology have proposed the concept of estuarine quality paradox (Elliott and Quintino 2007, Dauvin and Ruellet 2009), which demonstrates the challenge to detect the effects of anthropic factors on ecological status due to the variety of conditions that can occur in an estuary as a result of natural stressors. The benthic community structure found in our study followed the pattern described by Tommasi (1979), where the estuarine system was separated from the east and the west portions (including bay).

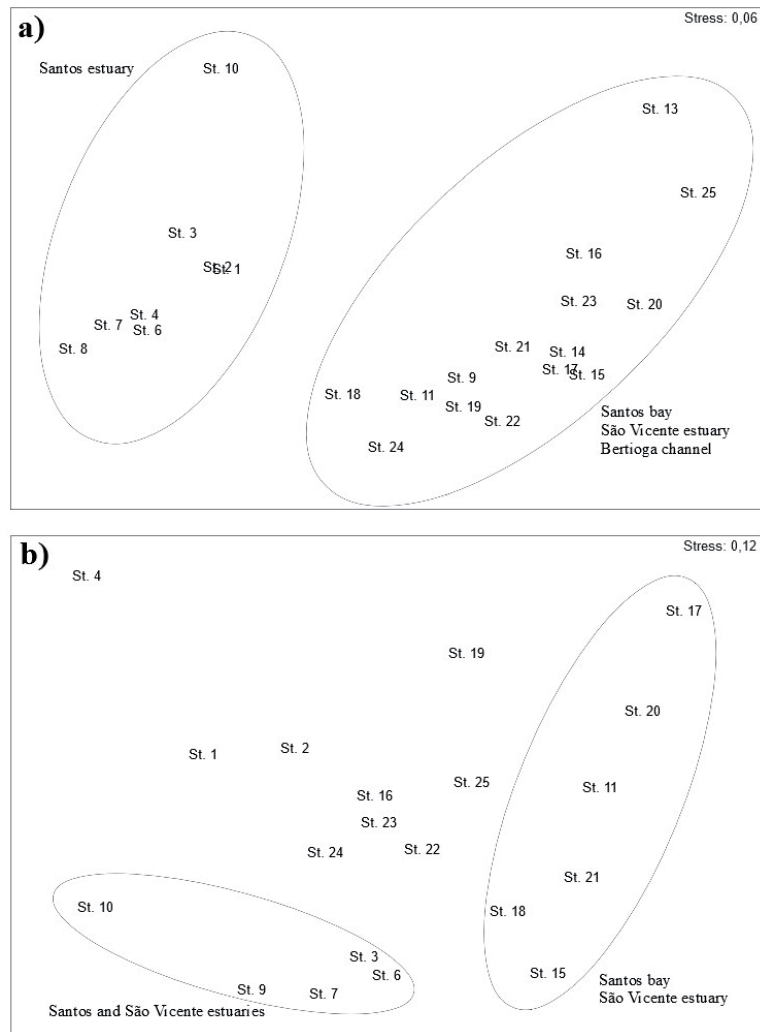


Figure 4. Ordination of samples based on nMDS results of environmental variables (a) and benthic community composition (b).

The author also pointed out that the existence of physiological stress gradients from the bay towards the estuary and in this case, such gradients were relevant for the most impacted sites (St 4 to 12). The convergence of both gradients of contamination and natural stressors in SES is an example of how the complexity of disturbances described on the estuarine quality paradox is relevant to describe benthic assemblages.

Data integration

To understand the influence of the natural stressors on the benthic community structure is a relevant issue and to explore that, both natural and anthropic factors were integrated with benthic fauna data, because we hypothesized that the combination of both is conditioning the structure of the communities in the SES. The first three components (PCs) of the PCA explained 68.51% variances (see Fig. 5 and Tables S9 to S12 in the Supplementary Material). The PC-1 represented 46.94% variances and accounted for positive correlations of water physical-chemical variables (temperature, pH, OD and salinity) and negative for the most

of contaminants (excepting MBAS) and sediment toxicity. Among species, *Semele purpurascens* correlated positively to the axis 1, while *Trichobranchus* sp., *Naineris setosa*., (Prob) *Pseudoerythoe* sp., *Pectinaria* sp., *Tetrasanthus rathbunae*, and *Kalliapseudes schubartii* presented negative correlations. These results suggest that species negatively correlated to the PC-1 were tolerant and/or favored by contamination, specifically at Santos Channel estuary (St. 1 to 9), São Vicente Channel (St. 11), SSOS diffusers (St. 18) and the inner portion of Bertioga Channel (St 24).

The PC-2 represented 14.27% of variances, with positive correlations found for temperature and Fe, and negative correlations reported for mud, CaCO₃, TOC, N and S, which characterizes the organic enrichment and the depositional areas. No species correlated positively to the PC-2, but *Marphysa* sp., *Glycera* sp.; Terebellidae (probably *Pista* sp.), *Ctena pectinella*, *Chione cancelata*, *Corbula patagonica* and *Tagellus* sp. were negatively correlated and they seem to be adapted to the such conditions, specifically at estuarine sites (St. 1 to #4, 9 and 10) and close to SSOS diffusers (St. 16, 18, 19 and 21). The PC-3 accounted for 7.296% variances and

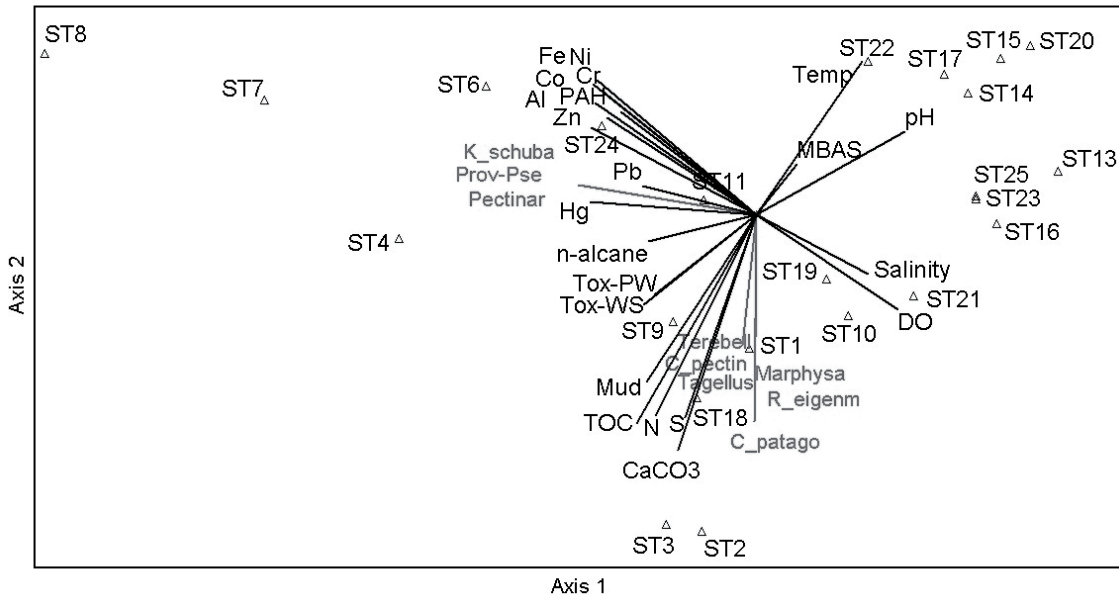


Figure 5. Results of the Principal Component Analysis ordination using data of geochemistry, toxicity, and benthic community obtained at the Santos Estuarine System.

presented positive correlations for n-alkanes and negative for temperature, S and MBAS. The polychaete *Magelona posterolongata* presented positive correlation to this PC, whereas the bivalve *Felaniella candeana* was negatively correlated.

The bi-dimensional ordination of axis 1 and 2 clustered one group composed by sites which exhibited organic enrichment, toxicity, and the presence of opportunistic species such as *Tagellus* sp. and *C. patagonia* (St.1, 2, 3, 9 and 18). The second group included sites within the Santos and Bertioga channels (St. 4, 6, 7, 8, 11 and 24), which sediments were contaminated by metals, n-alkanes, and PAHs; these sites also presented the occurrence of tolerant species such as *K. schubarti*, *Pseudoerythoe* sp. and *Pectinaria* sp. A third group separated low contaminated sites, located in the mouth of São Vicente and Bertioga channels (St 13 and 25, respectively), and Santos Bay (stations 14 to 17, 20, 22, and 23). The fourth group gathered stations influenced by marine conditions at Santos bay (St. 19 and 21) and a low number of organisms at São Vicente Channel (St 10) (Fig. 5).

The moderated amount of variance explained by the PCA reflects the heterogeneity of the studied environment, and indicates that other stressors may also influence the benthic composition of SES. Another issue is the elevated number of rare species, which resulted in a matrix composed of absent data. Other study carried out in Biscayne Bay, Florida, found a similar result of multivariate methods due to the complexity of conditions (Long et al. 2002). Van Dolah et al. (1999) pointed out that responses of benthic communities may be influenced by confounding factors, such as the number of absent species, making thus the integration of biological data with variables quite difficult. The alternative of using ecological indices may be also low efficient to detect environmental disturbances in these situations (Borja et al. 2000, Drake et al. 1999, Eaton 2001).

In summary, benthic assemblages were affected in sites from the inner portions of the SES and vicinities of the SSOS diffusers as Santos Bay (St 3 to 10, 18 and 24), which was supported by chemical contamination and sediment toxicity. Six stations exhibited moderately condition of disturbance (St. 1, 2, 11, 13, 22 and 17). In the stations St.1 and 2, the macrobenthic community was rich and diverse, and the concentrations of measured contaminants were low, but sediments were toxic. Other sites from the mouth of São Vicente and Bertioga channels (St. 14 and 25), and Santos Bay (St. 15, 16, 19, 20, 21, and 23) benthic fauna were considered not degraded. Recent studies conducted in this region produced similar results considering only ecological indices, but not the species composition (Buruaem et al. 2013; 2015). Thus, our results complement those findings by providing baseline information on the fauna composition at the time in different sectors of SES.

Conclusions

In this study, we described the existence of natural and anthropogenic stressors (i.e., grain size, contamination, and toxicity) in SES gradients that influence the structure of benthic communities. Benthic communities were more disturbed in sites influenced by contamination sources, as those listed by the Sao Paulo State Environmental Agency – CETESB (Lamparelli et al. 2001). The most critical area was the inner portion of Santos Channel (Piaçaguera Channel), due to the presence of an industrial complex. In the São Vicente Channel, despite the moderate contamination, the benthic community was also affected. In the Santos Bay, where natural factors seem to be more important in shaping the structure of communities, sites influenced by the SSOS were enriched in contaminants and organic materials, having additional effects on benthic organisms. The integration approach applied

in our study revealed that both natural and anthropogenic factors influenced the structure of benthic communities. The complexity of SES represents a challenge to evaluate its environmental quality status.

Acknowledgements: We would like to thank FAPESP (98/00808-6 Process) and PROPe-UNESP (Process 0067/017/13-PROPe/CDC) for the financial support; the IOUSP North Base staff for the logistic and general assistance; and the help of Professor L. Roberto Tommasi, Dr. F. Attolini, Professor E. Nonato (in memoriam), Dr. K. Christol dos Santos, Dr. M. T. Valério Belardo, Dr. M. F. Lopes dos Santos, Dr. A. M. Setúbal Pires Vanin and Dr. T. Navajas Corbisier, who kindly identified some taxonomic groups (respectively, echinoderms, polychaetes, tanaids, amphipods, cumaceans, isopods and anthozoans). We greatly appreciate the constructive technical reviews provided by Drs. M. Nipper, S. Carr and E. Long. D. Abessa also thanks CNPq by the PQ fellowship (Grants #303620/2008-0; #311609/2014-7). L.B. Moreira also thanks FAPESP (grant number 2013/15482-0).

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Received April 27, 2018
 Revised March 22, 2019
 Accepted May 24, 2019

Supplementary material

Table S1. Geographical coordinates and depths of each sampling station in the Santos Estuarine System.

Table S2. Physical-chemical parameters of water collected in each sampling station at Santos Estuarine System.

Table S3. Concentrations of Aliphatic Hydrocarbons (AHs) in sediments from the Santos Estuarine System (in µg/g).

Table S4. Concentrations of Poly-Aromatic Hydrocarbons (PAHs) in sediments from the Santos Estuarine System (in $\mu\text{g/g}$).

Table S5. Physical chemical parameters of the overlying waters measured in the test-chambers during the whole sediment toxicity test with the amphipod *T. viscana*, using the sediments from the Santos Estuarine System.

Table S6. Physical chemical parameters of the pore water samples extracted from the sediments from the Santos Estuarine System and tested with embryos of the sea-urchin *L. variegatus*.

Table S7. Presence or absence of toxicity and estimations of the NH_3 contents in pore water samples extracted from sediment collected in SES (T = toxic; NT = not toxic)

Table S8. Benthic species and their mean densities (in organisms per square meter) in the Santos Estuarine System.

Table S9. Variances extracted and Eigenvalues from the PCA with two matrices, using data of the Santos Estuarine System.

Table S10. First 3 Eigenvectors, each scaled to its standard deviation (correlation coefficients between scores for rows in the main matrix and the column variables), from the PCA ordination using data of the Santos Estuarine System. Values with bold fonts indicate significant correlations to the respective axes.

Table S11. Coordinates (Scores) of stations, according to the PCA run with two data matrices (data of the Santos Estuarine System).

Table S12. Pearson and Kendall Correlations with Ordination Axes (N= 23). Gray cells are showing significant correlations to the respective axes.

The Appendix may be downloaded from www.akademaii.com.