#### ORIGINAL ARTICLE

**Clinical and Experimental Dental Research** 

WILEY

## Thermal damage and excision time of micro and super pulsed diode lasers: A comparative ex vivo analysis

Mariliza C. O. Prado<sup>1</sup> | Ngozi N. Nwizu<sup>2</sup> | Shalizeh A. Patel<sup>3</sup> | Charles F. Streckfus<sup>2</sup>

<sup>1</sup>Laboratory of Biophotonics, Center for Lasers and Applications, IPEN-CNEN/SP, São Paulo, São Paulo, Brazil

<sup>2</sup>Department of Diagnostic and Biomedical Sciences, University of Texas Health Science Center at Houston, School of Dentistry, Houston, Texas, USA

<sup>3</sup>Department of Restorative Dentistry & Prosthodontics, University of Texas Health Science Center at Houston, School of Dentistry, Houston, Texas, USA

#### Correspondence

Juliana Barros, Department of Restorative Dentistry & Prosthodontics at the University of Texas Health Science Center at Houston, School of Dentistry; 7500 Cambridge St. suite 5350, 77054 Houston, TX, USA, Email: Juliana.Barros@uth.tmc.edu

Funding information

Biolase

# Denise Maria Zezell<sup>1</sup> I Juliana Barros<sup>3</sup>

#### Abstract

Objectives: The primary aim of this ex vivo study was to evaluate thermal damage and cutting efficiency of micro and super pulsed diode lasers. The secondary aim was to suggest a guideline to perform simple surgical excisions adequate for histopathological evaluation.

Material and Methods: Ten groups of 10 specimens of pig tongues were excised using a blade (G1), a micro pulsed (G2-G9), and a super pulsed diode (G10) lasers. Different output power, pulse duration, pulse interval, and duty cycle were tested. Quantitative measures of thermal damage and excision times were recorded. Statistical analysis was performed at a significance level of 5%.

Results: The control group (G1) presented no thermal damage. Within the laser groups (G2–G10), no statistically significant differences in depth of thermal damage  $(\mu m)$  were noted. G3 showed significantly less area of thermal damage  $(mm^2)$  when compared with G7 and G9 (p < .05). The median excision time of the control group and super pulsed diode laser group were significantly lower (p < .001) than the micro pulsed diode laser groups.

**Conclusions:** The cutting efficiency of the super pulsed diode laser is comparable to traditional blade, and with appropriate parameters, these lasers can produce predictable surgical outcomes with less collateral damage.

#### KEYWORDS

micro-pulsed diode laser, oral biopsy, super-pulsed diode laser, thermal damage

### 1 | INTRODUCTION

As standard of care, general dentists are to identify oral lesions, develop differential diagnoses, and provide appropriate treatment to patients. Currently, diode lasers (810-1064 nm) are used to perform various procedures such as ablation, incision, excision, and coagulation for intraoral surgeries. Dental diode lasers are minimally invasive and can become valuable in the treatment of soft tissue surgeries including biopsies. Hemostasis of surgical field, postoperative pain management, and faster healing rates are a few advantages of diode lasers well documented in the literature (Diamanti et al., 2002; Gontijo et al., 2005; Margarone et al., 1984; Wlodawsky & Strauss, 2004). As with any highintensity laser, diode lasers convert light into thermal energy. This interaction with the target tissue can produce different biological effects,

\_\_\_\_\_ This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2022 The Authors. Clinical and Experimental Dental Research published by John Wiley & Sons Ltd.

ranging from coagulation to carbonization (Kende et al., 2011). Diode lasers' unique affinity to interact with pigments within the targeted oral soft tissues, allows it to become a selective surgical tool. This leads to more controlled and less invasive surgical outcomes, reducing inflammation and pain and improving healing (Gold & Vilardi, 1994; Pick & Colvard, 1993; White et al., 1998).

However, laser-tissue interaction produces some degree of tissue vaporization including a zone of thermal damage around the incision site. Thus, without the use of the appropriate parameters, diode lasers can induce excessive heat damage in the excised tissue, interfering with its histopathologic evaluation and ultimate diagnosis (Gold & Vilardi, 1994; Kende et al., 2011; Pick & Colvard, 1993; White et al., 1998; Wilder-Smith et al., 1997).

High-intensity lasers can emit light in two modes: continuous wave and pulsed. Diode lasers, in particular, have a constant emission of radiation. In the newer diode devices, a mechanical shutter gates the light transmission, shortening the pulse width allowing for thermal relaxation of the target tissue (Coluzzi & Convissar, 2011; Zezell et al., 2011). This feature allows the power to be delivered in repetitive pulse, ranging from milliseconds (ms) to microseconds (µs). It is important to note that a laser with a shorter pulse duration usually results in a lower temperature and thereby produces less carbonization of the tissue (Hobbs et al., 1987). The newest diode lasers are now equipped with super and micro pulsed technology, controlling its duty cycle, the fraction of time the laser is actively emitting energy (expressed in a percentage). This feature allows the laser to deliver high power in a short period of time, protecting the tissue from severe heat damage. It would be beneficial to carry out an ex vivo study to confirm such claims. Therefore, the primary aim of this ex vivo study was to evaluate the effects of different parameters on the integrity of collected specimens and the cutting efficiency of the two lasers' pulse modalities. The secondary aim was to suggest a general guideline for clinicians to perform simple surgical excisions adequate for histopathological evaluation using diode lasers.

#### MATERIAL AND METHODS 2

This study was IRB exempted; however, all the principles of safety, ethical, and professional conduct have been followed by the authors, including specimen disposal. Informed consent was not required for this study. Fresh pig tongues (within 24 h of slaughter) were used in this ex vivo study. The pig tongues were stored at a temperature of 2°C-4°C to prevent tissue degradation before the experiment (Gomez-Santos et al., 2010). All excisions were made at room temperature between 22°C and 24°C (Melo et al., 2010). A 940 nm diode laser (Epic X<sup>™</sup> - micro pulsed diode laser and Epic Pro<sup>™</sup> - super pulsed diode laser, Biolase, Irvine, CA, USA) with 300 µm and 400 µm disposable-initiated surgical tips, respectively, were used. One hundred standardized specimens with 8 mm diameter were excised from the dorsal portion of pig tongues (Figures 1 and 2) and randomly distributed into 10 groups (n = 10) as shown in Table 1. The control group (G1) biopsies were performed with conventional technique using 15C a scalpel blade. The specimens assigned to the micro pulsed diode laser groups (G2-G9) were excised





FIGURE 1 Picture of the pig tongue receiving a standardized 8 mm diameter delineation before laser excision.



FIGURE 2 Picture of the excisional biopsy procedure on a pig tongue using a 300-µm initiated fiber.

at two average powers of 1.0 W and 1.5 W (Azma & Safavi, 2013; Gutiérrez-Corrales et al., 2020; Romeo et al., 2014). Other variables within the laser unit were emission mode (continuous vs. pulsed), pulse duration (10 µs, 100 µs, and 1 ms), pulse interval (40 µs, 200 µs, and 1 ms), and duty cycle (20%, 33%, and 50%). The specimens assigned to the super pulsed diode laser group (G10) were removed at an average power of 3.2 W, peak power of 80 W, super pulse width of 10 µs-10 ms, pulse interval 0.01 ms-20 ms, and pulse repetition rate ranging up to 20 kHz (Romanos et al., 2016).

The output power was checked before the start of each surgical procedure using a power meter (PM600 Power Meter, Molectron Detector Inc, Portland, OR, USA). The surgical time of each excision was recorded from the time of the laser application on the tissue to the complete removal of the sample. The specimens were fixed in 10% buffered formalin solution, cut into three equal-sized slices, and embedded in paraffin blocks, according to conventional histological processing methods (Feldman & Wolfe, 2014; Fox et al., 1985). Serial sectioning of 4 µm thickness was performed for subsequent staining with hematoxylin and eosin (H&E). The histopathological images were evaluated under vertical light microscopy (Eclipse 80i, Nikon Corporation, Tokyo, Japan) and were captured with Andor Zyla 5.5 sCMOS (Andor Technology Ltd. Company, Belfast, UK). Each slide was

	Group	Pulse mode	Duty cycle	Pulse Repetition rate (Hz)	Power at display (W)	Power measured (W)	Power density calculated (W/cm <sup>2</sup> )	Peak power at display (W)	Pulse width	Pulse Interval (off time)
Scalpel	G1 ( <i>n</i> = 10)	Control	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a
Epic X <sup>TM</sup> - micro pulsed diode laser (940 nm)	G2 (n = 10)	CW	n/a	20,000	1.2	1	1415.4	1.2	n/a	n/a
	G3 ( <i>n</i> = 10)			20,000	1.8	1.5	2123.1	1.8		
	G4 ( <i>n</i> = 10)	СРО	20%	20,000	1.2	1	1415.4	Ŷ	10 µs	40µs
	G5 (n = 10)			20,000	1.8	1.5	2123.1	6		
	G6 ( <i>n</i> = 10)	CP1	33%	20,000	1.2	1	1415.4	3.6	100 µs	200 µs
	G7 (n = 10)			20,000	1.8	1.5	2123.1	5.4		
	G8 ( <i>n</i> = 10)	CP2	50%	20,000	1.2	1	1415.4	2.4	1 ms	1 ms
	G9 (n = 10)			20,000	1.8	1.5	2123.1	3.6		
Epic Pro <sup>m</sup> - super pulsed diode laser (940 nm)	G10 ( <i>n</i> = 10)	STP	n/a	40	n/a	3.2	2547.7	80	10 μs-100 ms	0.01 ms-20 ms

**TABLE 1** Group distribution and diode lasers parameters

1657

WILEY



**FIGURE 3** Representative images of the two thermal damage measurements (area and depth). (a) Measurement of the total area of damage is delineated in blue and calculated in millimeter square (mm<sup>2</sup>). (b) Measurement of depth of thermal damage is outlined in yellow in micrometers ( $\mu$ m).

observed at 40X magnification using the Nikon Eclipse 80i Advanced Research Microscope software (Nikon Instruments Inc., Melville, NY, USA). Both area (mm<sup>2</sup>) and depth ( $\mu$ m) of tissue thermal damage were recorded. The total area of thermal damage was quantified by calculating the difference between the total area of each specimen and its total area of damage, visually detected by the microscope (Figure 3a). Next, the area with the most thermal damage on each specimen was identified. A virtual 90° line from the base of the specimen was drawn within the most heataffected area (Figure 3b).

Statistical analyzes were performed using the IBM<sup>®</sup> SPSS software package, V20.0 (IBM Corp., Armonk, NY, USA) with a significance level of 5%. Shapiro–Wilk normality test was performed. The observed distributions were asymmetric and described by medians and quartiles, minimum and maximum values. Kruskal–Wallis nonparametric tests were applied to compare the study groups in relation to thermal damage areas, thermal damage depths, and excision times. Differences were reported using Dunn's multiple comparison tests with Bonferroni correction. The correlation between depth of thermal damage and percentage of area with thermal damage was evaluated by Spearman's rank correlation test.

#### 3 | RESULTS

#### 3.1 | Area of thermal damage

The control group (G1) presented no thermal damage. In experimental groups (G2-G10), the observed total area of

TABLE 2	Total area of thermal damage (mm <sup>2</sup> ) of the specimens
according to	the study group

Group	Minimum	First quartile	Median	Third quartile	Maximum	n
G2	0.46	1.03	1.09	1.54	1.98	10
G3	0.40	0.53	0.91*	1.11	2.60	10
G4	0.71	0.91	1.22	1.86	2.99	10
G5	0.93	1.26	1.37	1.66	3.93	10
G6	0.34	1.20	1.53	1.72	1.90	10
G7	1.32	1.52	1.93*	2.45	3.71	10
G8	0.73	1.09	1.50	2.42	3.37	9
G9	0.77	1.40	1.97*	3.50	3.97	10
G10	0.57	0.90	1.08	1.74	2.76	9

\*Kruskal-Wallis test: p = .009. G3 was significantly different when compared to G7 and G9; G1, the control group was excluded from the thermal damage evaluation as no laser was used.

thermal damage (mm<sup>2</sup>) was lower in the G3 group (1.5 W, CW) with a median of  $0.91 \text{ mm}^2$  and p < .05 (Table 2). Applying multiple comparison tests and controlling at the global significance level, the median of the damaged area of the G3 and G10 groups were lower than the adjusted medians of G7 (1.5 W, 33% duty cycle) and G9 (1.5 W, 50% duty cycle) groups. No statistically significant differences were observed in the other groups.

#### 3.2 | Depth of thermal damage

The control group (G1) presented no thermal damage. In experimental groups (G2–G10), the depth of thermal damage ( $\mu$ m) in the specimens indicated no evidence of statistical differences amongst the groups (*p* = .125) (Table 3).

## 3.3 | Correlation of area and depth of thermal damage

The total area of thermal damage (mm<sup>2</sup>) ranges from 0.91 to 1.97 mm<sup>2</sup> (Table 2) and the depth of thermal damage ranges from 606 to 1024  $\mu$ m (Table 3). A positive correlation was noted between the depth ( $\mu$ m) and area (mm<sup>2</sup>) when evaluating thermal damage (Figure 4).

#### 3.4 | Excision time

It is noteworthy that the median excision time of the super pulsed diode laser group (G10) is comparable to the control group

Group	Minimum	First quartile	Median	Third quartile	Maximum	n
G2	317.6	489.4	606.4	858.1	1114.7	10
G3	468.0	574.5	655.8	742.8	863.2	10
G4	462.0	566.8	720.4	779.9	918.3	10
G5	429.7	482.5	789.6	1080.0	1206.2	10
G6	465.1	590.8	669.3	960.4	1129.8	10
G7	599.5	839.8	1024.9	1393.5	1555.0	10
G8	437.3	510.5	666.4	849.7	1397.9	9
G9	319.0	584.6	831.1	1057.5	1414.0	10
G10	470.6	530.5	778.8	947.1	1203.6	10

*Note*: Kruskal–Wallis test: p = .125. No statistical significance was detected amongst all groups. G1, the control group was excluded from the thermal damage evaluation as no laser was used.

(G1-scalpel). Excision time medians in G1 and G10 were significantly lower (p < .001) than the medians of the micro pulsed laser groups (G2-G9) (Table 4).

### 4 | DISCUSSION

Several studies have demonstrated that CO<sub>2</sub>, Er:YAG, Nd:YAG, and diode lasers can be used in oral soft tissue biopsies without causing significant thermal damage and hindering the histopathological diagnosis (Fox et al., 1985; Merigo et al., 2013; Romeo et al., 2014). However, very few have evaluated and compared variables such as thermal damage and excision times produced by micro and super pulsed diode lasers, the most common surgical lasers in clinical dentistry (Angiero et al., 2012; Belikov et al., 2015; Fornaini et al., 2016; Monteiro et al., 2019; Romeo et al., 2014). To bridge this gap, the present study evaluated two diode lasers with different combinations of power, duty cycle, and pulse duration. The findings of this study support the view that different parameters can variously affect soft tissue responses, such as thermal damage and cutting efficiency. In addition, this study showed a positive correlation

#### TABLE 4 Excision time(s) of specimen according to study group

Group	Minimum	First quartile	Median	Third quartile	Maximum	n
G1	38	42	50*	68	108	10
G2	187	201	238	270	350	10
G3	122	179	194	227	244	10
G4	97	147	197	213	317	10
G5	110	122	161	194	258	10
G6	116	130	155	190	293	10
G7	131	142	155	158	306	9
G8	106	138	193	248	309	9
G9	110	122	142	150	205	9
G10	62	66	69*	75	83	10

\*Kruskal–Wallis test: p < .001. Note that G1 (scalpel) and G10 (super pulsed diode laser) were statistically significantly different from all the other groups (p < .05).



**FIGURE 4** This graph illustrates the positive correlation between depth ( $\mu$ m) and area (mm<sup>2</sup>) caused by thermal damage.

between area and depth of tissue affected when measuring overall thermal damage. Thus, our discussion reflects findings on both methodologies, when referencing thermal damage.

This present study noted that the micro pulsed diode laser G3 (CW, 1.5 W) produced less total area of thermal damage (mm<sup>2</sup>) when compared to all groups. The groups with the greatest area of thermal damage (mm<sup>2</sup>) were G9 (1.5 W, 50% duty cycle) and G7 (1.5 W, 33% duty cycle), while the least damage was identified as G3 (CW, 1.5 W) followed by G10 (super pulsed laser, 3.2 W). These observations can partially be explained by a phenomenon called the "hot tip" effect in combination with the laser's pulse duration. The hot tip effect is caused when a target tissue absorbs the laser energy, leading to protein denaturation and carbonization of the tissue. This leads to a carbonized build-up at the tip of the fiber, which continues to overheat, into several hundred degrees, and promotes tissue cutting, coagulation, and possible thermal damage (Verdaasdonk & Swol, 1997). Therefore, it is recommended to constantly stop and remove this carbonized build-up debris from the laser tip. Though removing this aggregate of debris is needed, the cleaned tip has now a reduced ability to superficially interact with the tissue. This allows the laser light to penetrate deeper into the target tissue. As this carbonization build-up and cleaning cycle continues, a fluctuation in the depth of thermal penetration and cutting efficiency within the same surgical site is noted.

In addition to the variance in this heat penetration, the laser pulse duration is another major factor that can negatively impact tissue response to uncontrolled thermal damage. Laser pulse duration is referred to the time tissue is exposed to laser energy. The longer the time, the more laser-tissue interaction is observed (Belikov et al., 2015: Spencer et al., 1998). Additionally, the amount of power (peak vs. average) can dictate the degree of interaction within the surgical site. Peak power is the maximum optical power a laser can produce to interact with the target tissue within a specific time frame. Average power is the power transferred to the target tissue per unit of time. Regardless of what power parameter is utilized, it is important to balance thermal effects within a tissue by allowing the thermal energy to dissipate. This time of cooling is referred to as thermal relaxation (Stuart Nelson et al., 1995). A proper laser protocol must encourage the reduction of heat-generated collateral damage by balancing both power and pulse duration. Pulse duration greater than 1µs has displayed noticeable thermal effects on oral soft tissue (Niemz, 2007). In the present study, groups G7 and G9 with larger pulse duration (100  $\mu$ s and 1 ms) and a high peak power (5.4 W and 3.6 W) resulted in a greater area of heat transmission within the specimens. In contrast, the super pulse diode laser (G10) had a pulse duration longer than  $1 \mu s$  (10  $\mu s$ -10 ms) and high peak power (80 W), but caused less thermal penetration. This can be explained by the new tip technology within this super pulsed diode laser (Epic Pro™), where a controlled thermal feedback mechanism is installed to read tissue's temperature and adjust its level at the fiber tip during excision time. This advanced system aims to prevent overheating of the tissue and improve cutting efficiency (Romanos et al., 2016). Our study findings showed good congruence with those of Wilder-Smith

et al. (1997); they observed that incisions created with a super pulsed mode  $CO_2$  laser (pulse duration of 300 µs, peak power 60–100 W, and average power of 0.7–1.2 W) showed a significant reduction in collateral thermal damage when compared with continuous mode (Wilder-Smith et al., 1997). Our results also corroborated those of Romanos' findings, as the time duration for removing biopsied tissues obtained with the super pulsed diode laser (G10), was significantly faster than other groups (Romanos et al., 2016). In addition, it was noted that the cutting efficiency of the super pulsed diode laser was comparable to a scalpel.

The present study utilized pig tongues as they are histologically and physiologically similar to human tongues (Azevedo et al., 2016; Cercadillo-Ibarguren et al., 2010; Romeo et al., 2010). The investigators selected an 8 mm diameter as the standardized specimen size for two main reasons. Laser surgery is not recommended for any lesion less than 5 mm in size due to the high probability of heat-induced artifacts interfering with microscopical evaluation (Angiero et al., 2012; Vescovi et al., 2010). Second, 8 mm represents the average size of most simple excisional biopsies of common oral lesions +1 mm allowance of a halo of healthy peripheral tissue for proper histological analysis. Besides, the larger specimen size is easier to handle in an ex-vivo setting. For example, fibromas which are the most frequent (53.3%) benign tumors of the oral cavity vary in size between 5 and 8 mm (Allon et al., 2014; Gonsalves et al., 2007; Torres-Domingo et al., 2008). The most common locations for these slowly growing lesions are the buccal mucosa along the occlusal line and the tongue (Gonsalves et al., 2007; Torres-Domingo et al., 2008). Treatment is surgical excision and recurrence is rare. Per literature, it is advised when using laser as a surgical tool, to leave a halo of 0.5-1 mm of healthy tissue peripheral to pathology. This preserves the morphological and structural characteristics of the collected specimen allowing for a readable and reliable diagnosis (Romeo et al., 2014). According to this study, the deepest area of thermal damage within the specimen ranged in on average from 606 to 1024 µm supporting Romeo et al suggestion of leaving a band of healthy tissue peripheral to diseased margins to compensate for any heatinduced artifacts (Romeo et al., 2010).

To the best of the authors' knowledge, this is the first study to evaluate the effects of different parameters of micro and super pulsed diode lasers on the integrity of tissue specimen and its cutting efficiency. Therefore, it provides invaluable baseline information that would be useful for further studies. Diode lasers are the most common surgical lasers utilized by general dentists, thus their proper use, including appropriate biopsy parameters should be standardized. This study provides useful information that can guide clinicians in the laser excision of simple surgical biopsies, improve surgical outcomes and patient safety.

For study limitations, the use of pig tongue mucosa, despite the similarities, is not human tongue mucosa, the authors recognize there is a possibility that there might be some variations in the results obtained using actual human tongue mucosa. Furthermore, this study had several limitations regarding the comparison between the new

-Wii fy

1661

20574347, 2022. 6, Downloaded from https://onlinelibrary.wiley.com/doi/10.1002/cre2.670 by Univ of Sao Paulo - Brazil, Wiley Online Library on [23/12/2022]. See the Terms

and Conditions

(https://onl:

.wiley

ons) on Wiley Online Library for rules of use; OA articles are governed by the applicable Creative Commons License

super pulsed laser and the micro pulsed laser, since in the latter it is not possible to control the peak power and pulse width, as it presents an intrinsic variable, unlike the super pulsed laser. Another limitation of this study is the attainment of an exact specimen size in a reproducible manner. Even though a diameter of 8 mm was set, all measurements were not always exactly precise because laser cutting of the tissue was done manually and may have introduced the possibility of small random errors in mm<sup>2</sup>. To overcome this limitation, however, the investigators tested two methods of measuring thermal damage (depth and area). After statistical analysis, it was concluded that both measurements correlated positively and can be applied independently or collectively.

Future clinical studies are needed to confirm this study's finding of high peak power and long pulse duration of micro pulsed laser can generate greater thermal damage in comparison to super pulsed lasers.

#### CONCLUSION 5

In this study, the investigators aimed to (1) evaluate the thermal effects of different parameters on the integrity of collected specimens excised by micro and super pulsed diode lasers, (2) compare the cutting efficiency of the two diode laser systems, and (3) suggest a general guideline for clinicians to perform simple surgical excisions adequate for histopathological evaluation. The results of the study confirmed the use of diode lasers as a suitable surgical tool for excision of simple benign tumors when appropriate parameters are used. A minimum of 1 mm perimeter of healthy margins is recommended for appropriate microscopic analysis of the tissue specimen. This "safe zone" will be sufficient to withstand collateral damage caused by laser heat under the parameters described in this study. In addition, the super pulsed diode laser with the new thermal feedback system demonstrated a more efficient cutting ability than the micro pulsed laser and was comparable to the scalpel. However, not many clinicians are able to replace their existing lasers with the latest technology as they become available. Thus, it is relevant to apply knowledge gained from this ex vivo study to suggest general guidelines for best clinical outcomes. Per results of this study, clinicians using:

- conventional diode lasers may consider a continuous mode of emission rather than a 50% duty cycle;
- micro pulsed diode laser may select the shorter pulsed duration parameter offered within the laser or continuous mode of emission:
- super pulsed diode laser may select the lowest cutting power parameter offered within this new generation of lasers.

The above guideline can reduce heat-generated collateral damage allowing for proper histopathological evaluation of biopsied tissues. Nevertheless, there is a need for further randomized clinical trials comparing these types of diode lasers to precisely establish their differences and to recommend parameters for the best results in a clinical setting.

#### AUTHOR CONTRIBUTIONS

Dr. Mariliza Prado has contributed to the material preparation, data collection, and analysis, original draft of the manuscript. Dr. Ngozi Nwizu has contributed to the study conception and design, data analysis, review, and editing of the manuscript. Dr. Shalizeh Patel has contributed to the material preparation, data analysis, review, and editing of the manuscript. Dr. Charles Streckfus has contributed to the statistical analysis and review of the manuscript. Dr. Denise Zezell has contributed to the study conception and design, review, and editing of the manuscript. Dr. Juliana Barros has contributed to the study conception and design, material preparation, data analysis, review, and editing of the manuscript. All authors read and approved the final manuscript.

#### ACKNOWLEDGMENTS

The authors wish to acknowledge the following colleagues for their contribution to this project Ms. Connie Dieringer, Ms. Candy Bales from UT Health School of Dentistry (Houston), and Sandra Regina Malagutti from Sao Paulo - SP, Brazil.

#### CONFLICTS OF INTEREST

The authors have received a grant for laser consumable supplies from Biolase, Inc. Irvine, USA that partially supported this study.

#### DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

#### **ETHICS STATEMENT**

This study does not contain any involving human participants or animals. For this type of ex vivo study, no ethical committee was needed, the current study is in accordance with the ethical standards of The University of Texas Health Science Center at Houston.

#### ORCID

Denise Maria Zezell D https://orcid.org/0000-0001-7404-9606 Juliana Barros D https://orcid.org/0000-0003-0472-4949

#### REFERENCES

- Allon, I., Kaplan, I., Gal, G., Chaushu, G., & Allon, D. (2014). The clinical characteristics of benign oral mucosal tumors. Medicina Oral Patología Oral y Cirugia Bucal, 19(5), e438-e443.
- Angiero, F., Parma, L., Crippa, R., & Benedicenti, S. (2012). Diode laser (808 nm) applied to oral soft tissue lesions: A retrospective study to assess histopathological diagnosis and evaluate physical damage. Lasers in Medical Science, 27(2), 383-388. https://doi.org/10.1007/ s10103-011-0900-7
- Azevedo, A., Monteiro, L., Ferreira, F., Delgado, M., Garces, F., Carreira, S., Martins, M., & Suarez-Quintanilla, J. (2016). In vitro histological evaluation of the surgical margins made by different laser wavelengths in tongue tissues. Journal of Clinical and Experimental Dentistry, 8(4):e388-e396. https://doi.org/10.4317/jced.52830

- Azma, E., & Safavi, N. (2013). Diode laser application in soft tissue oral surgery. *Journal of Lasers in Medical Sciences*, 4(4), 206–211.
- Belikov, A. V., Skrypnik, A. V., & Shatilova, K. V. (2015). Comparison of diode laser in soft tissue surgery using continuous wave and pulsed modes in vitro. Frontiers of Optoelectronics, 8(2), 212–219. https:// doi.org/10.1007/s12200-015-0517-8
- Cercadillo-Ibarguren, I., Espana-Tost, A., Arnabat-Dominguez, J., Valmaseda-Castellon, E., Berini-Aytes, L., & Gay-Escoda, C. (2010). Histologic evaluation of thermal damage produced on soft tissues by CO<sub>2</sub>, Er,Cr:YSGG and diode lasers. *Medicina Oral Patología Oral y Cirugia Bucal*, 15(6), e912–e918. https://doi.org/10.4317/medoral. 15.e912
- Coluzzi, D. J., & Convissar, R. A. (2011). Laser fundamentals. In Convissar, R. A. (Ed.). *Principles and Practice of laser dentistry* (pp. 12–26). Mosby.
- Diamanti, N., Duxbury, A. J., Ariyaratnam, S., & Macfarlane, T. V. (2002). Attitudes to biopsy procedures in general dental practice. British Dental Journal, 192(10), 588–592. https://doi.org/10.1038/sj.bdj. 4801434
- Feldman, A. T., & Wolfe, D. (2014). Tissue processing and hematoxylin and eosin staining. Methods in Molecuar Biology, 1180, 31–43. https:// doi.org/10.1007/978-1-4939-1050-2\_3
- Fornaini, C., Merigo, E., Sozzi, M., Rocca, J. P., Poli, F., Selleri, S., Cucinotta, A. (2016). Four different diode lasers comparison on soft tissues surgery: a preliminary. *Laser Therapy*, 25(2), 105–114. https://doi.org/10.5978/islsm.16-OR-08
- Fox, C. H., Johnson, F. B., Whiting, J., & Roller, P. P. (1985). Formaldehyde fixation. Journal of Histochemistry & Cytochemistry, 33(8), 845–853. https://doi.org/10.1177/33.8.3894502
- Gold, S. I., & Vilardi, M. A. (1994). Pulsed laser beam effects on gingiva. Journal of Clinical Periodontology, 21(6), 391–396. https://doi.org/10. 1111/j.1600-051x.1994.tb00735.x
- Gomez-Santos, L., Arnabat-Dominguez, J., Sierra-Rebolledo, A., & Gay-Escoda, C. (2010). Thermal increment due to ErCr:YSGG and CO<sub>2</sub> laser irradiation of different implant surfaces. A pilot study. *Medicina Oral Patología Oral y Cirugia Bucal*, 15(5), e782–e787. https://doi. org/10.4317/medoral.15.e782
- Gonsalves, W. C., Chi, A. C., & Neville, B. W. (2007). Common oral lesions: Part I. Superficial mucosal lesions. *American Family Physician*, 75(4), 501–507.
- Gontijo, I., Navarro, R. S., Haypek, P., Ciamponi, A. L., & Haddad, A. E. (2005). The applications of diode and Er:YAG lasers in labial frenectomy in infant patients. *Journal of Dentistry for Children* (*Chicago*, III), 72(1), 10–15.
- Gutiérrez-Corrales, A., Rizcala-Orlando, Y., Montero-Miralles, P., Volland, G., Gutiérrez-Pérez, J., Torres-Lagares, D., & Serrera-Figallo, M. (2020). Comparison of diode laser - Oral tissue interaction to different wavelengths. In vitro study of porcine periodontal pockets and oral mucosa. *Medicina Oral Patología Oral y Cirugia Bucal*, 25(2), e224–e232. https://doi.org/10.4317/ medoral.23317
- Hobbs, E. R., Bailin, P. L., Wheeland, R. G., & Ratz, J. L. (1987). Superpulsed lasers: minimizing thermal damage with short duration, high irradiance pulses. *The Journal of Dermatologic Surgery and Oncology*, 13(9), 955–964. https://doi.org/10.1111/ j.1524-4725.1987.tb00571.x
- Kende, P. R. G., Yuwanati, M., & Jain, B. (2011). Application of diode laser in oral biopsy: Removal of White patch over tongue -A case report. Journal of the Indian Dental Association, 5, 958-987.
- Margarone, J., Natiella, J. R., & Natiella, R. (1984). Primates as a teaching model for biopsy. *Journal of Dental Education*, 48(10), 568-570.

- Melo, M. R., Martins, A. R., Barbosa, I. V., Romano, P., & Shcolnik, W. (2010). Coleta, transporte e armazenamento de amostras para diagnóstico molecular. Jornal Brasileiro de Patologia e Medicina Laboratorial, 46(5), 375–381. https://doi.org/10.1590/S1676-2444 2010000500006
- Merigo, E., Clini, F., Fornaini, C., Oppici, A., Paties, C., Zangrandi, A., Fontana M., Rocca J.P., Meleti M., Manfredi M., Cella L., Vescovi P. (2013). Laser-assisted surgery with different wavelengths: A preliminary ex vivo study on thermal increase and histological evaluation. *Lasers in Medical Science*, 28(2), 497–504. https://doi. org/10.1007/s10103-012-1081-8
- Monteiro, L., Delgado, M., Garces, F., Machado, M., Ferreira, F., Martins, M., Salazar, F., & Pacheco, J. (2019). A histological evaluation of the surgical margins from human oral fibrousepithelial lesions excised with CO2 laser, diode laser, Er:YAG laser, Nd:YAG laser, electrosurgical scalpel and cold scalpel. *Medicina Oral Patología Oral y Cirugia Bucal*, 24(2), e271–e280. https://doi.org/10. 4317/medoral.22819
- Niemz, M. H. (2007). Laser-Tissue Interactions: Fundamentals and Applications (3rd ed.). Springer.
- Pick, R. M., & Colvard, M. D. (1993). Current status of lasers in soft tissue dental surgery. *Journal of Periodontology*, 64(7), 589–602. https:// doi.org/10.1902/jop.1993.64.7.589
- Romanos, G. E., Altshuter, G., & Yaroslavsky, I. (2016). Epic Pro Reinventing Diode Laser Soft - Tissue Therapy using Science and Technology. BIOLASE.
- Romeo, U., Palaia, G., Del Vecchio, A., Tenore, G., Gambarini, G., Gutknecht, N., De Luca, M. (2010). Effects of KTP laser on oral soft tissues. An in vitro study. *Lasers in Medical Science*, 25(4), 539–543. https://doi.org/10.1007/s10103-010-0756-2
- Romeo, U., Russo, C., Palaia, G, Lo Giudice, R., Del Vecchio, A., Visca, P., Migliau, G., De Biase A. (2014). Biopsy of different oral soft tissues lesions by KTP and diode laser: Histological evaluation. *The Scientific World Journal*, 2014, 1–6. https://doi.org/10.1155/2014/761704
- Spencer, P., Cobb, C. M., Wieliczka, D. M., Glaros, A. G., & Morris, P. J. (1998). Change in temperature of subjacent bone during soft tissue laser ablation. *Journal of Periodontology*, 69(11), 1278–1282. https:// doi.org/10.1902/jop.1998.69.11.1278
- Stuart Nelson, J., Milner, T. E., Svaasand, L. O., & Kimel, S. (1995). Laser pulse duration must match the estimated thermal relaxation time for successful photothermolysis of blood vessels. *Lasers in Medical Science*, 10, 9–12. https://doi.org/10.1007/BF02133157
- Torres-Domingo, S., Bagan, J. V., Jiménez, Y., Poveda, R., Murillo, J., Díaz, J. M, Sanchis J.M., Gavaldá C., Carbonell E. (2008). Benign tumors of the oral mucosa: a study of 300 patients. *Medicina Oral*, *Patologia Oral y Cirugia Bucal*, 13(3), 161–166.
- Verdaasdonk, R. M., & Swol, C.F.P. (1997). Laser light delivery systems for medical applications. *Physics in Medicine and Biology*, 42(5), 869–894. https://doi.org/10.1088/0031-9155/42/5/010
- Vescovi, P., Corcione, L., Meleti, M., Merigo, E., Fornaini, C., Manfredi, M., Bonanini, M., Govoni P., Rocca J.P., Nammour S. (2010). Nd:YAG laser versus traditional scalpel. A preliminary histological analysis of specimens from the human oral mucosa. *Lasers in Medical Science*, 25(5), 685-691. https://doi.org/10. 1007/s10103-010-0770-4
- White, J. M., Chaudhry, S. I., Kudler, J. J., Sekandari, N., Schoelch, M. L., & Silverman S, J.r (1998). Nd:YAG and CO<sub>2</sub> laser therapy of oral mucosal lesions. *Journal of Clinical Laser Medicine & Surgery*, 16(6), 299–304. https://doi.org/10.1089/clm.1998.16.299
- Wilder-Smith, P., Dang, J., & Kurosaki, T. (1997). Investigating the range of surgical effects on soft tissue produced by a carbon dioxide laser. The Journal of the American Dental Association, 128(5), 583-588. https://doi.org/10.14219/jada. archive.1997.0257

WILEY

- Wlodawsky, R. N., & Strauss, R. A. (2004). Intraoral laser surgery. Oral and maxillofacial surgery clinics of North America, 16(2), 149–163. https:// doi.org/10.1016/j.coms.2004.02.001
- Zezell, D. M., Ana, P. A., Pereira, T. M., Correa, P. R., & Velloso, W. J. (2011). Heat generation and transfer on biological tissues due to high-intensity laser irradiation. In Bernardes, M. A. S. (Ed.). *Developments in heat transfer* (Chap. 13, pp. 227–246). InTech. Accessed January 2022. https://www. intechopen.com/books/developments-in-heat-transfer/heat-generationand-transfer-on-biological-tissues-due-to-high-intensity-laser-irradiation

How to cite this article: Prado, M. C. O., Nwizu, N. N., Patel, S. A., Streckfus, C. F., Zezell, D. M., & Barros, J. (2022). Thermal damage and excision time of micro and super pulsed diode lasers: A comparative ex vivo analysis. *Clinical and Experimental Dental Research*, 8, 1655–1663. https://doi.org/10.1002/cre2.670