PREPARATION OF SAMARIUM-153-EDTMP AND DETERMINATION OF ITS RADIOCHEMICAL PURITY USING PAPER CHROMATOGRAPHY

Haroldo Taurian Gasiglia, Helena Okada

Comissão Nacional de Energia Nuclear/SP, Instituto de Pesquisas Energéticas e Nucleares, Caixa Postal 11049, Pinheiros, 05422-970, São Paùlo/SP, Brazil

> Received 12 October 1994 Accepted 26 October 1994

Samarium-153-ethylenediaminetetramethylene-phosphonate (153 Sm-EDTMP) has been used to palliate pain resulting from bone cancer. This paper describes the preparation of 153 SmCl $_3$ from irradiated natural samarium and the ability of 153 Sm to complex with EDTMP in liquid and in freeze-dried forms. The evaluation of radiolabeled EDTMP was done by paper chromatography. A rapid evaluation of free 153 SmCl $_3$ and 153 Sm-EDTMP was developed using a miniaturized chromatographic system.

INTRODUCTION

Recent advances in targeted radiotherapy offer a new approach for management of metastatic bone pain. Interest in employing radiopharmaceuticals for diagnosis and therapy has increased and significant advances have been pos-

sible by combined developments in the fields of new chelates and availability of a wider range $\beta\text{-emitting}$ nuclides 1 .

Samarium-153 has excellent physical properties as radiotherapeutic agent. It is a β -emitter (E $_{\beta max}$ = 0.8 meV) with 46.8 h half-life. The average β -particle energy is approximately one-third of its maximum energy. Samarium-153 also emits a 103-keV γ -ray suitable for imaging with conventional γ -cameras 2 , 3 . This radioisotope is produced by neutron activation of $^{152}{\rm Sm}$. When $^{152}{\rm Sm}$ (99% enriched $^{152}{\rm Sm}$) is irradiated in high thermal fluxes the $^{153}{\rm Sm}$ yields are in the order of curies. The radioactive impurities ($^{154}{\rm Eu}$, $^{155}{\rm Eu}$) appear in trace amounts 2 .

Samarium-152 thermal cross-section is 220 barns, but it also has an epithermal resonance integral cross-section of 3,168 barns that will increase the production in reactors with high epithermal fluxes³.

Reactors with thermal fluxes of about 1x10¹³ n.cm⁻². sec⁻¹ can produce ¹⁵³Sm from natural samarium (26.7% ¹⁵²Sm) in the order of millicuries. These yields are suitable for in vitro studies and for distribution studies in mice. Radioactive impurities like ¹⁵²Eu, ¹⁵⁴Eu and ¹⁵⁵Eu obtained when natural samarium is irradiated do not affect these studies.

Samarium forms complexes with several phosphonic acids 2,4 . Among $^{153}{\rm Sm}$ chelates, $^{153}{\rm Sm}$ -EDTMP has shown to be a promising agent for the treatment of bone cancer.

When ¹⁵³Sm is chelated to EDTMP, it forms stable complexes in vitro and in vivo, which will localize in bone with a high specificity². The structure of ethylenediaminetetramethylenephosphonic acid (EDTMP) is depicted in Figure 1.

Fig. 1. Structure of EDTMP

Samarium-153-EDTMP complex can be prepared from EDTMP in solution under heating 2,4,5 or from kit formulation, at room temperature 6,7 .

This work describes the ¹⁵³Sm-EDTMP labeling, the yields obtained in both methods and the stability of ¹⁵³Sm-EDTMP using paper chromatography.

EXPERIMENTAL

Reagents: Natural samarium oxide, BDH, analytical grade.

EDTMP, Monsanto Chemical Corporation. Other

reagents: Merck, analytical grade. pH strips:

Merck Neutralit.

Equipments: 153 Sm activities: Radioisotope Calibrator Capintec model CRC 120. Chromatogram strips countings: ANSR Gamma Counter, Abbot Laboratories. EDTMP titrations: pH-meter Metrohm Herisau Model E-520.

Preparation of 153 SmCl₃

Natural samarium oxide was converted into samarium nitrate by dissolution with 1N nitric acid. Samarium

amounts of 2 to 4.1 mg were put into a high purity quartz vial and evaporated to dryness, leaving a thin film of samarium nitrate on the botton of the vial. It was flame-sealed and encapsulated into an aluminium container.

The samples were irradiated in the research reactor IEAR-1. Its maximum available thermal flux is 1.5×10^{13} n.cm⁻².sec⁻¹

After irradiation, the quartz vial was removed from its aluminium container in a hot cell. The sample was measured for $^{153}\mathrm{Sm}$ activity and transferred to a glove box. The quartz vial was broken at the top and 2x1 ml of 0.1N hydrochloric acid were introduced. The vial was heated (80-90 $^{\mathrm{O}}\mathrm{C}$) and the $^{153}\mathrm{Sm}$ solution was transferred to a glass vial. $^{153}\mathrm{SmCl}_3$ solutions were diluted with 0.1N hydrochloric acid to produce a stock solution (0.9 mg Sm ml $^{-1}$) used for complex preparations.

Preparation of 153sm-EDTMP

The 153 Sm-EDTMP complex was prepared as follows:

- a) EDTMP in solution: The 153 Sm-EDTMP was prepared by adding 1.0 ml of 153 SmCl $_3$ solution (0.9 mg Sm) to 60 mg/1.5 ml EDTMP at pH 9 (molar ratio 1:23, Sm/EDTMP). The pH of reaction mixture was raised to 10-11 with sodium hydroxide solution. The mixture was placed into a 60-70 $^{\circ}$ C water bath for 30 min. After heating the pH was adjusted to 7.5-8 with hydrochloric acid.
- b) EDTMP in lyophilized form: A kit formulation consists of 60 mg EDTMP titrated with sodium hydroxide to pH 10.3 and further lyophilized. Addition of 1.7 ml of a 153 SmCl₃ solution in 0.1N hydrochloric acid (0.9 mg Sm) to the kit yields a solution with pH of 7.5-8 (molar ratio 1:23, Sm/EDTMP).

Radiochemical purity of ¹⁵³sm-EDTMP

Three chromatography systems were tested for ¹⁵³Sm-EDTMP radiochemical purity determination.

System 1: Support: Whatman 3MM paper strips 1.5x13.0 cm. Solvent: pyridine, ethanol, water 1:2:4 v/v/v.

System 2: Support: Whatman 3MM paper strips 1.5x13.0 cm. Solvent: ammonium hydroxide, ethanol, water 0.1:2:4 v/v/v.

System 3: Support: Whatman 3MM paper strips 1.0x7.0 cm (miniaturized).

Solvent: ammonium hydroxyde, ethanol, water 0.1:2:4 v/v/v.

The chromatograms were developed for a distance of 11.0 cm in approximately 75-85 min (systems 1 and 2) and for a distance of 5.0 cm in 20-25 min (system 3).

For $R_{\rm f}$ determinations the strips were cut in 1 cm (system 1 and 2) and 0.5 cm (system 3) pieces and the activity of each piece was measured. The radioactivity was assumed to be concentrated at the center of each piece.

Studies of 153 Sm-EDTMP stability

The stability of $^{153}\text{Sm-EDTMP}$ obtained in both preparations was investigated for 8 d.

RESULTS AND DISCUSSION

Yields of ¹⁵³Sm

A run of 7 to 8 h produced the $^{153}\mathrm{Sm}$ yields listed in Table 1. The quartz vials measured after $^{153}\mathrm{Sm}$ transference to the glass vials showed negligible residual activities.

GASIGLIA, OKADA: PREPARATION OF 153Sm-EDTMP

TABLE 1 $$^{153}\rm{Sm}$$ yields obtained at a thermal flux of $1.5 \times 10^{13} n.\, \rm cm^{-2}.\, sec^{-1}$ at the end of irradiation

Sample No.	Samarium mass, mg	Irradiation time,	Activity, mCi Theoretical Obtained		
1	2.5	7.56	25.4	19.9	
2	3.1	8.00	33.1	24.6	
3	4.1	7.55	43.7	33.6	
4	2.0	7.45	20.0	18.6	
5	3.1	7.00	29.1	24.3	
6	2.0	7.70	20.6	19.1	
7	4.1	8.20	44.7	33.7	
8	3.0	8.13	32.5	25.7	

Chromatographic behavior of 153 SmCl3 and 153 Sm-EDTMP

Figure 2 shows the chromatographic behavior of $^{153}\mathrm{Sm}$ -EDTMP compared with that of $^{153}\mathrm{SmCl}_3$ using systems 1 and 2. Figure 3 shows the chromatographic behavior using system 3.

The ranges of $\rm R_{f}$ values found for $^{153}\rm SmCl_{3}$ and $^{153}\rm Sm-EDTMP$ are presented in Table 2.

For a rapid and easy estimate of the radiochemical purity of ¹⁵³Sm-EDTMP, after setting up the radioactive distribution (system 3), the strips were cut in two portions. Then the activity of each portion was determined. In Figure 3 the position for strips cutting is indicated. This procedure was used for all complexing yield determinations.

Complexing yields

In basic medium $^{753} \text{Sm-EDTMP}$ can be readily prepared with complexing yields higher than 97.5%.

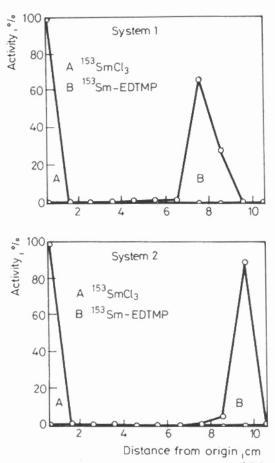


Fig. 2. Chromatographic behavior of $^{153}\mathrm{Sm-EDTMP}$ and $^{153}\mathrm{SmCl}_3$ using systems 1 and 2

The complexing yields in both methods (n=6) are given in Table 3.

¹⁵³Sm-EDTMP stability

The complex remained stable for at least 8 d after labeling using EDTMP in solution and after reconstitution of the kits with $^{153}\rm{Sm/HCl}$. Table 4 summarizes the complex stability over time.

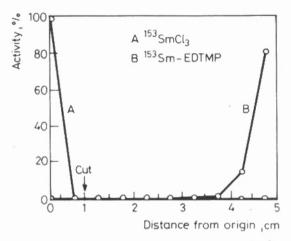


Fig. 3. Chromatographic behavior of $^{153}\mathrm{Sm-EDTMP}$ and $^{153}\mathrm{SmCl}_3$ using system 3

TABLE 2 ${\rm Range\ of\ R_{f}\ values\ for\ }^{153}{\rm SmCl}_{3}\ {\rm and\ }^{153}{\rm Sm-EDTMP}$

System	153 _{SmCl₃}	153 _{Sm-EDTMP}	
1	0	0.70-0.75	
2	0	0.85-0.90	
3	0	0.88-0.94	

ADDENDUM

Further studies showed that the developing time in the miniaturized chromatographic system can be reduced to approximately 15 min. This is possible when the solvent mixture ammonium hydroxide, ethanol, water 0.1:2:4, v/v/v is replaced by a solvent mixture ammonium hydroxide, methanol, water 0.2:2:4 v/v/v. No significant alterations

GASIGLIA, OKADA: PREPARATION OF 153Sm-EDTMP

TABLE 3
%Complex yields of ¹⁵³Sm-EDTMP (1st day)

Labeling No.	In solution	EDTMP In kit formulation
1	98.8	98.6
2	98.4	98.2
3	99.1	99.2
4	99.2	97.9
5	99.0	98.5
6	98.3	98.8
Mean label	ing 98.8±0.4	98.5±0.5

TABLE 4 153 Sm-EDTMP stability (%complexing yield)

Day	Labeling EDTMP in solution			Number EDTMP in kit formulation		
	1	3	6	1	3	6
1	98.8	99.1	98.3	98.6	99.2	98.8
2	98.1	98.4	99.0	98.3	99.7	98.2
4	98.2	98.5	98.7	98.6	99.5	98.5
8	98.1	99.0	98.2	98.7	99.1	98.4

were observed for $\rm R_f$ values of either $^{153}\rm SmCl_3$ and $^{153}\rm Sm-EDTMP$ or for EDTMP complexing yields.

We wish to thank Dr. Mitiko Saiki, from IPEN/CNEN-SP for $\rm Sm_2^O{}_3$ supply, M.Sc HoWoui Ling Wang from IPEN/CNEN-SP for her help in preparating this manuscript and for Monsanto do Brasil for EDTMP supply.

GASIGLIA, OKADA: PREPARATION OF 153Sm-EDTMP

REFERENCES

- 1. A.J.B. McEwan, Eur. J. Nucl. Med., 20 (1993) 1.
- W.F. Goeckeler, D.E. Troutner, W.A. Volkert, B. Edwards, J. Simon, D.W. Wilson, <u>Nucl. Med. Biol.</u>, 13 (1986) 479.
- W.A. Volkert, W.F. Goeckeler, G.J. Ehrhardt, A.R. Ketring, J. Nucl. Med., 32 (1991) 174.
- W.F. Goeckeler, B. Edwards, W.A. Volkert, R.A. Holmes, J. Simon, D. Wilson, J. Nucl. Med., 28 (1987) 495.
- Sh.Zh. Luo, M.F. Pu, J. Quao, Zh.L. Liu, Ch.Y. Zhang, P.J. Zhao, Y.B. Fu, H.F. Deng, <u>J. Radioanal. Nucl.</u> Chem. Articles, 160 (1992) 443.
- J.R. Garlich, S.A. Baugman, J. Simon, K. McMillan, A.R. Ketring, W.A. Volkert, W.F. Goeckeler, D.E. Trouther, J. Label. Compds, Radiopharm., XXIII (1986) 1341.
- A. Singh, R.A. Holmes, M. Farhangi, W.A. Volkert, A. Williams, L.M. Stringham, A.R. Ketring, <u>J. Nucl. Med.</u>, 30 (1989) 1814.